



**MARLBOROUGH
DISTRICT COUNCIL**

Soil Quality in the Marlborough Region in 2017

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Executive Summary

Regional councils (and unitary councils) have a responsibility for promoting the sustainable management of the natural and physical resources of their region. Under Section 35 of the Resource Management Act (1991), one of the physical resources that we have a duty to monitor and report on is soil. Specifically, to report on the “life supporting capacity of soil” and to determine whether current practices will meet the “foreseeable needs of future generations”. To help meet these goals, the Marlborough District Council (Council) undertakes a soil quality monitoring programme that involves collecting soil samples from a network of sites that represent the main land use activities and soil types within the region and analysing these samples for a suite of soil physical, biological and chemical properties that have been shown to be robust indicators of soil quality. The aim of this report is to summarise both the current state of, and the long-term trends in, soil quality in the Marlborough region as determined by the results of soil analysis from sampling across a range of land use activities and soil types.

In this investigation, soils were sampled from twenty four monitoring sites that included 8 vineyards, 5 dairy sites, four pasture sites, four exotic forests, one cropping site and two under native lowland forest. These sites represented 11 different soil types from 4 soil orders.

This year’s results are similar to last year’s results. While many sites show good soil quality, most soils show the effects of human land use with soil quality indicators for many of these falling outside target ranges. 65% of sites (mostly vineyards) reported soil compaction measurements outside the target range. These results put these soils at risk of poor aeration and impeded drainage which may potentially affect pasture production and predispose the soil to surface runoff, nutrient loss, erosion and flooding. While soil compaction may not be permanent, it clearly should be avoided and remediated where necessary. A range of beneficial management options to prevent and remediate soil compaction are outlined in the report.

Cadmium levels remain a concern for dairy farmers but rising levels in other farmed land uses should not be ignored, especially in the context that 65% of this year’s sites reported low phosphate levels. Cadmium is a contaminant of phosphate fertiliser, additional fertiliser will be required to maximise production on properties with low P values. While levels are not likely to exceed target ranges in the short-term, continued use of phosphate fertiliser will mean levels will continue to rise and continued monitoring will be required

The long-term analysis introduced in 2016 has been repeated this year. The results from a new set of sites confirm the concerns outlined in last year’s report that soil compaction, soil organic matter loss and loss of nutrients to water are significant problems for Marlborough.

A new section has been introduced this year looking at landscape change. Changes from pastoral production to viticulture are commonplace in Marlborough. The results show major reductions in soil organic matter and increases in soil compaction occurring within 12 months of vineyard conversion. It should be emphasised that this analysis is based on only 2 sites and very small data sets. If the data is accurate, it implies large amounts of carbon are being lost from soils.

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1. Introduction

Regional councils have a responsibility for promoting the sustainable management of the natural and physical resources of their region. Under Section 35 of the Resource Management Act (1991), one of the physical resources that we have a duty to monitor and report on is soil. Specifically, to report on the “life supporting capacity of soil” and to determine whether current practices will meet the “foreseeable needs of future generations”. The collection of detailed soil monitoring data is therefore vital because it provides information on what effect current land use activities are having on soil quality and whether we need to change or prioritise the way we manage the land environment. This is becoming increasingly important as land use activities such as dairying and viticulture are intensifying across New Zealand and putting pressure on our soils.

Furthermore, the way soils respond to different land use activities can affect other parts of the environment, for example water quality. This is because soils act as buffers to capture and store nutrients such as nitrogen, phosphorous and microbes, treat a range of waste products and store and filter water.

To help determine what effect land use practices are having on soil quality, in 2000 the Marlborough District Council (MDC) became a participant in a national soil quality monitoring programme known as “The 500 Soils Project”. At the completion of this project the MDC implemented its own soil quality monitoring programme commencing in 2008 to continue assessing the quality of soils throughout the Marlborough region. This programme is largely based around the framework developed as part of the national programme and is in line with soil quality monitoring currently undertaken in other regions in New Zealand.

The objectives of the soil quality monitoring programme are to:

- Provide information on the physical, chemical and biological properties of soils to assess overall soil health;
- Provide an early-warning system to identify the effects of primary land uses on long-term soil productivity and the environment;
- Track specific, identified issues relating to the effects of land use on long term soil productivity;
- Assist in the detection of spatial and temporal changes in soil quality; and
- Provide a mechanism to determine the effectiveness of regional policies and plans.

A network of 96 soil quality monitoring sites has been established in Marlborough. This report presents results for; 15 sites previously sampled in 2000, 2007 and 2012. 5 sites previously sampled in 2008 and 2013 as well as 2 sites last sampled in 2009 and 2014. The wider spread of sites reflects changes made last year to the frequency of sampling to help improve data reliability. Three sites (27, 28 &9) have been sampled specifically to capture soil quality changes that may have occurred following conversion to vineyard. The report discusses if they meet their target ranges for soil quality.

2. Materials and Methods

2.1. Sampling Sites

Soils were sampled from; 15 sites previously sampled in 2000, 2007 and 2012, 5 sites previously sampled in 2008 and 2013 as well as 2 sites last sampled in 2009 and 2014. They include 8 vineyard sites, two sites under native lowland forest, five dryland pasture sites, four exotic forestry sites, two dairy sites and one cropping site. Some sites scheduled for sampling this year were not sampled in order to improve temporal spread of sites, nine sites were brought forward for the same reason. The sampled sites represent 11 different soil types from 4 soil orders (Table 1).

Table 1 Soil type, soil classification and land use management of sites sampled in Marlborough

Site Code	Sampling years	Soil Type	Soil Order*	Land use; management
SOE_Soils_Site01	2000, 2007, 2012, 2017	Wairau	Recent	Vineyard
SOE_Soils_Site02	2000, 2007, 2012, 2017	Wairau	Recent	Vineyard
SOE_Soils_Site04	2000, 2007, 2012, 2017	Renwick	Pallic	Vineyard
SOE_Soils_Site05	2000, 2007, 2012, 2017	Renwick	Recent	Cropping
SOE_Soils_Site06	2000, 2007, 2012, 2017	Renwick	Pallic	Dryland Pasture
SOE_Soils_Site07	2000, 2007, 2012, 2017	Paynter	Pallic	Vineyard
SOE_Soils_Site08	2000, 2007, 2012, 2017	Paynter	Gley	Vineyard
SOE_Soils_Site09	2000, 2007, 2012, 2017	Paynter	Pallic	Vineyard- <i>Recent conversion</i>
SOE_Soils_Site14	2000, 2007, 2012, 2017	Seddon	Pallic	Dryland Pasture
SOE_Soils_Site17	2000, 2007, 2012, 2017	Kaituna	Recent	Native Bush
SOE_Soils_Site19	2000, 2007, 2012, 2017	Kenepuru	Brown	Native Bush
SOE_Soils_Site20	2000, 2007, 2012, 2017	Kenepuru	Brown	Exotic Forest
SOE_Soils_Site21	2000, 2007, 2012, 2017	Kenepuru	Brown	Exotic Forest
SOE_Soils_Site22	2000, 2007, 2012, 2017	Kenepuru	Brown	Dryland Pasture
SOE_Soils_Site24	2000, 2007, 2012, 2017	Wairau	Recent	Dryland Pasture
SOE_Soils_Site27	2008, 2013, 2017	Motukarara	Gley	Vineyard
SOE_Soils_Site28	2008, 2013, 2017	Motukarara	Gley	Vineyard- <i>Recent conversion</i>
SOE_Soils_Site40	2008, 2013, 2017	Kaituna	Brown	Dairy
SOE_Soils_Site41	2008, 2013, 2017	Rai	Recent	Dairy
SOE_Soils_Site42	2008, 2013, 2017	Pelorus Steepland	Brown	Exotic Forest
SOE_Soils_Site43	2008, 2013, 2017	Pelorus Steepland	Brown	Dairy
SOE_Soils_Site44	2008, 2013, 2017	Koromiko	Brown	Dairy
SOE_Soils_Site52	2009, 2014, 2017	Tuamarina	Brown	Dryland Pasture
SOE_Soils_Site53	2009, 2014, 2017	Tuamarina	Brown	Exotic forest

*New Zealand Soil Classification

2.2. Soil Sampling

Three types of soil samples are collected from each site. Firstly, a composite sample comprising 25 individual cores taken at 2 m intervals along a 50 m transect at a depth of 100 mm (Plate 1a). These samples were combined into one large sample and used for chemical and organic matter analysis. A second set of 5 cores taken along the same transect at 10m intervals (10, 20, 30, 40, 50m) were collected and individually bagged to provide samples for a separate microbial ecology study. In addition, three undisturbed soil cores (100 mm diameter by 75 mm depth) were sampled at 15-, 30- and 45-m positions along the transect (Plate 1b). These soil cores were removed as one unit by excavation around the liner, bagged and loaded into padded crates for transport to the laboratory for analysis. These soil samples were used for soil physical analysis. Samples were collected from mid-October to early November. Most sites had reasonable soil moisture conditions.



Plate 1 (a) Collecting a composite of core samples along a transect using a soil corer. (b) One of three intact core samples taken at each site, to establish the physical properties of the soil.

2.2.1. Changes to sampling sites

The location of sampling sites should not change. However, a key objective of this project is to monitor land use and landscape changes to these sites. The majority of the sites sampled in this round are being sampled for the second or third time. This means some sites are now up to 17 years old and may have changed markedly from the original. Field notes from past sampling rounds help staff to locate the original transects so samples can be replicated as closely as possible. However, it has not been possible to replicate exactly the location of the original transect on some sites. Reasons for this include large changes in vegetation (especially in forested areas), errors in GPS location markers and unclear field notes. Where transects could not be located accurately, a new transect was established as closely as possible to the original using the original site photographs. New transects were documented with explicit notes and photographs to ensure location in the future. New transects were established on Sites 04 (vineyard redevelopment), 17 (repeat of erroneous sample from 2016), 19, 20, 21 (difficult to locate forest transects), & 28 (vineyard conversion) this year.

2.2.2. Viticulture sampling sites

Because of the economic importance and scale of viticulture in Marlborough, it was decided in 2012 that vineyard monitoring should encompass 3 samples per vineyard site. Samples are taken from under the vines, in the wheel tracks and in the inter-row region. This is done to allow the impact of various management practices to be evaluated. These include:

- Under vine
 - banding of fertiliser
 - herbicide applications
 - maintenance of bare ground
 - absence of traffic
 - irrigation
 - transfer of inter-row mowings
- Wheel tracks
 - soil compaction
- Inter-row
 - inputs of organic matter including prunings
 - lower rates of fertiliser
 - rainfall inputs only

2.3. Soil Quality Measurements

A number of different soil properties are measured to assess soil quality. Soil chemical characteristics were assessed by soil pH, Olsen P and trace element concentrations. Soil organic matter characteristics are determined by measuring anaerobically mineralisable nitrogen, total carbon, total nitrogen and carbon: nitrogen ratio.

Soil physical conditions were assessed using bulk density, particle density and water release characteristics which in turn were used to calculate total soil porosity, air filled porosity and macroporosity (Table 2). Additional microbial samples are being analysed to provide insight into soil ecological properties. This work will be reported on later as results become available.

Table 2 Indicators used for soil quality assessment

Indicators	Soil Quality Information	Method
Chemical properties		
Soil pH	Acidity or alkalinity	Glass electrode pH meter,
Olsen P	Plant available phosphate	Bicarbonate extraction, molybdenum blue method
Trace elements	Deficiency or toxicity of trace elements in soil	Acid digestion
Biological properties		
Anaerobically mineralisable N	Readily mineralisable nitrogen reserves	Waterlogged incubation at 40 °C for 7 days
Total Carbon	Organic matter status	Dry combustion, CNS analyser
Total Nitrogen	Organic N reserves	Dry combustion, CNS analyser

Indicators	Soil Quality Information	Method
Carbon: Nitrogen Ratio	Decomposition rate of organic matter	Calculated from above
Physical properties		
Dry bulk density	Compaction, volumetric conversions	Soil cores
Particle density	Used to calculate porosity and available water	Specific gravity
Total porosity, air capacity and macroporosity	Soil compaction, aeration, drainage	Pressure plates

2.4. Soil Analysis

Descriptions of the different soil analysis process are detailed below. In general, analysis follows the processes described by (Hill & Sparling, 2009) for soil quality parameters and (Kim & Taylor, 2009) for trace element analysis.

2.4.1. Chemical Analysis

All chemical analysis was undertaken by Hills Laboratory, Hamilton. Soil pH was measured in a 1:2 (v/v) soil:water slurry followed by potentiometric determination of pH (Blakemore, 1987). Soil phosphorus is determined with Olsen extraction followed by Molybdenum Blue colorimetry (Olsen, Cole, Watanabe, & Dean, 1954). Trace element determination made by Nitric/hydrochloric digestion followed by ICP-OES (Hills Laboratories, 2018).

2.4.2. Organic Matter Analysis

Biological analysis was carried out by Hills Laboratory, Hamilton. Anaerobically mineralisable nitrogen was estimated Anaerobic incubation followed by extraction using 2M KCl followed by Berthelot colorimetry (Keeney & Bremner, 1966). Total carbon and nitrogen were determined by dry combustion of air-dry soil (Hills Laboratories, 2018).

2.4.3. Physical Analysis

Soil physical analysis was undertaken by Landcare Research in Hamilton. Dry bulk density was measured on soil samples extruded from cores and dried in an oven at 105°C until the weight remained constant and the sample was then weighed (Gradwell & Birrell, 1979). Air filled porosity (-10 kPa) and total porosity were calculated as described by Klute, (1986). Particle density was measured by the pipette method.

It is worth noting that the general definition of macroporosity has recently been expanded to cover a slightly larger range of pores sizes than the original definition. Several regional councils have adopted macroporosity measurements based on the volumetric water content at -10kPa (technically referred to as the air filled porosity). So, in this report for consistency with other regions we now use the -10kPa measurement (defined in this report as air filled porosity), although the -5kPa data is included for reference because this has been used and reported by the MDC and others in the past.

2.4.4. Targets and Ranges

To aid in the interpretation of soil quality indicators, an expert panel (in several workshops) developed guidelines for the seven soil quality indicators now commonly used by regional councils (Hill & Sparling, 2009). The panel determined target ranges for the assessment of soil quality (e.g. very low, optimal, very high etc.) for the predominant soil orders under different land uses. The interpretative ranges from Hill & Sparling, (2009) are presented in Appendix A. However, Olsen P targets have recently been revised with new target values reported in (Mackay, Dominati, & Taylor, 2013) and used in this report (Appendix A).

The trace element results (except for cadmium) have been compared against the soil limits presented in the New Zealand Water and Wastes Association (NZWWA, 2003) 'Guidelines for the Safe Application of Biosolids to Land in New Zealand' (referred to as the biosolids guidelines) (Appendix A). While guidelines containing soil contaminant values like the biosolids guidelines have been written for a specific activity (i.e. biosolids application), the values are generally transferable to other activities that share similar hazardous substances. Cadmium results were compared to values in the Tiered Fertiliser Management System (TFMS) from the New Zealand Cadmium Management Strategy (MAF, 2010).

2.4.5. Data Display and Analysis

Readers of previous Soil Quality reports will note a number of changes in the presentation of the data. Firstly, the names of the sites were changed in 2016 in order to provide better referencing in the Council computer database. Sites were previously labelled using an "MDC" number e.g. MDC 15. These have now been renamed *SOE_Soils_Site15*. The number of each site remains the same. Vineyard sites are labelled *SOE_Soils_Site63a_vine, b_wheel* or *c_inter-row*.

The second change in data presentation from previous reports has been to present data in groups according to soil order or land use. This change allows the reader to more clearly understand how a soil conforms to its target values which are set according to soil order or land use. These two factors have the greatest influence on soil quality. Readers can refer to Appendix A for target ranges of soil quality indicators. Information on the New Zealand Soil Classification can be found at <https://soils.landcareresearch.co.nz/describing-soils/nzsc>

This report displays data in two ways. Firstly, Tables 3 & 4 shows the raw chemical and biological data from the bulked sample. Table 5 shows the physical data for each sample as a single averaged value for the three soil cores extracted from each site. Secondly, the long-term data uses a 5-year rolling average. Each data point for a given year is the average of all data from the proceeding 5 years for that data. This is done to smooth data outliers and to help illustrate long-term trends in the data.

As a result of the on-going review process for the Soil Quality Monitoring program, it has been identified that some land uses have insufficient sites to justify presenting this data as annual values (5 –year rolling average). Readers of the 2016 report will have noted the inconsistencies in native forestry results. This is due to the low number of sites (five) that are available for sampling (a reflection of the small number of remaining lowland native forests in Marlborough). Previous procedure was to sample these on a 10-year rotation; however, this has resulted in only 11 sample points for this land use over the entire course of the programme. Exotic forestry follows a similar pattern although this land use has 17 total sample points. As a result, data from these land uses is no longer presented as a time series but as a single point derived from the average of all data for that land use. This provides a reference for these land uses while reducing the effect of individual outlier data points. Future options for this data is to identify more sites or sample more often although costs limitations may constrain this.

As mentioned in section 2.4.3, previous Soil Quality reports used the term Macroporosity to refer to soil pore measurements. However, these measurements used data more correctly called air filled porosity. This report now refers to air filled porosity -10kPa to clarify which measurement is being used.

This report discusses changes in soil quality indicators over time. This is done to improve the understanding of soil quality changes on a regional basis. This has allowed the determination of some key issues for land managers to be aware of. See section 4 for further details.

Where appropriate, data were expressed on a weight/volume or volume/volume basis to allow comparison between soils with differing bulk density. Olsen P values are reported in different units (mg/L) than previous reports to account for differences in soil bulk density between samples.

3. Results and Discussion

3.1. Comparison of Target Ranges

Figure 1 shows the percentage of samples not meeting their target for a specific soil quality indicator. All samples for pH and total N met their target ranges. Olsen P, C: N ratio, and air filled porosity all showed a large number of samples failing to meet the target ranges (26, 14, 24 samples respectively). Trace elements, AMN, total C, and bulk density had smaller numbers not meeting their soil quality target. 40 samples were taken from 24 sites in 2017.

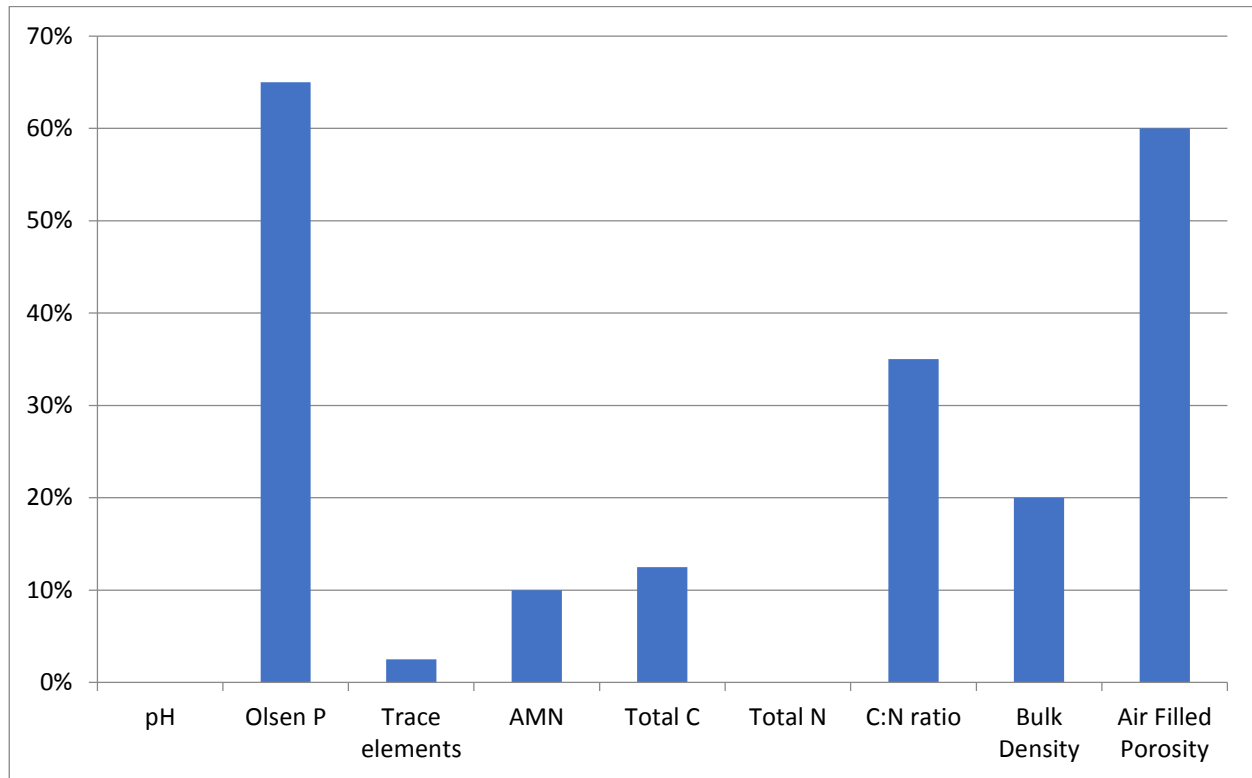


Figure 1: The percentage of sites not meeting their target range for a specific soil quality indicator

The results of soil chemical, biological and physical analyses from soils sampled at each site are given in Tables 3, 4 and 5 respectively and are discussed separately below.

3.2. Soil Chemical results

Results of soil chemical analysis (pH, Olsen P and Trace elements) are reported in Table 3. Each of the chemical properties is discussed individually. The target values appropriate to the relevant soil order can be found in Appendix A.

3.2.1. Soil pH

Soil pH is a measure of the acidity and alkalinity in soil. It is an important soil indicator because it affects nutrient and contaminant availability in plants and the functioning of beneficial soil macro- and micro-organisms. Most plants and soil organisms will have an optimum pH range for growth, and the pH of the soil affects which species will grow best.

As indicated in Table 3, all sites had soil pH values within the acceptable target for their respective land use. Differences are evident between land uses. Vineyards have slightly higher pH than other land uses with forestry (native and exotic) having the lowest pH (most acidic) readings. Analysis of pH by soil type shows no significant difference in pH between soil types.

Figure 2 is a box and whisker plot that shows the maximum and minimum values (whiskers), first and third quartiles (top and bottom of the box) and the median (central line in the box). The differences seen in Figure 2 are most likely due to land use. This is probably a reflection of fertiliser practice under the different land uses. Low input land uses will tend to lower pH due to the natural acidifying effects of plant growth. Farmed land will often receive fertiliser (and lime) relative to the value of the products coming from this land. As a result, it is common practice to apply fertiliser and lime annually to vineyards with a consequent lift in pH. The lower returns and larger-scale from pastoral farming often restrict fertiliser applications to limiting nutrients only. This seems to have led to an overall lower pH for pastoral land uses.

Interestingly, the average pH values shown for viticulture in 2017 are in the high end of the range that is normally preferred (5.8-6.8). This is similar to the longer-term samples (Figure 3). This would imply that some sites have much higher pH. Although it is not possible to determine with this data set, the implication is that pH management may need to be improved. Because of the regular application of fertiliser to vineyards, often small amounts of nutrients will be applied. This often requires lime to be added to the other nutrients in order for these to be spread effectively. Given the increased emphasis on sound nutrient management (and the financial costs of fertiliser), it is suggested that vineyard managers may wish to examine fertiliser practice more closely.

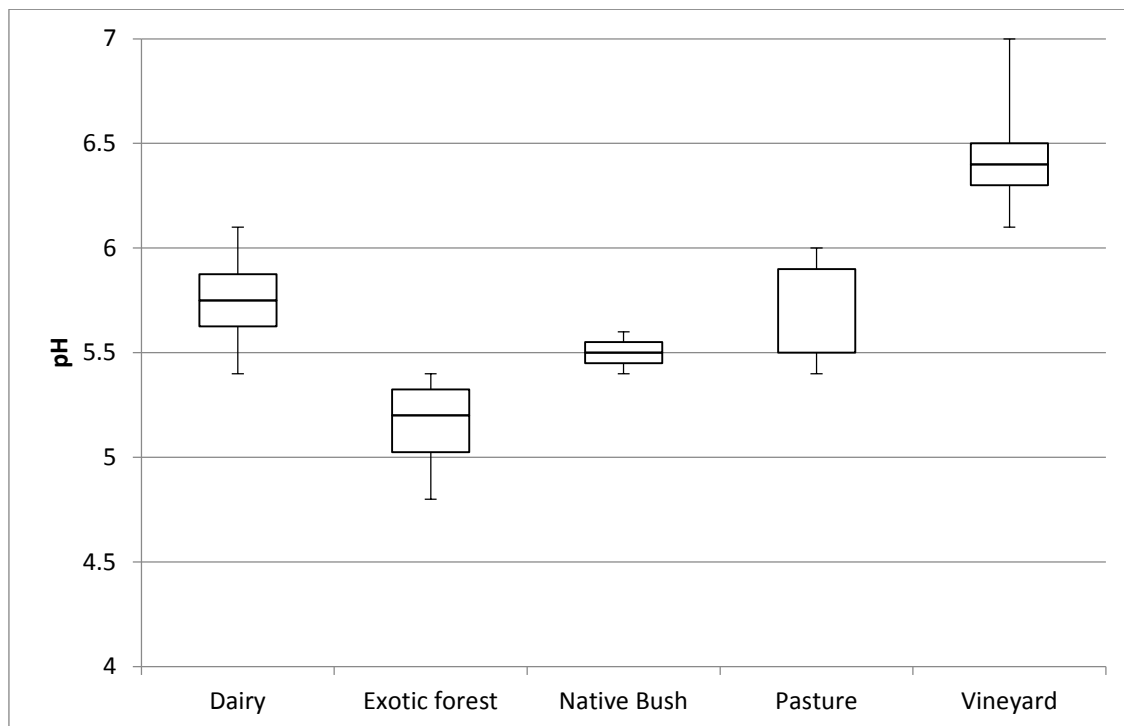


Figure 2: Soil pH by land use for 2017 samples

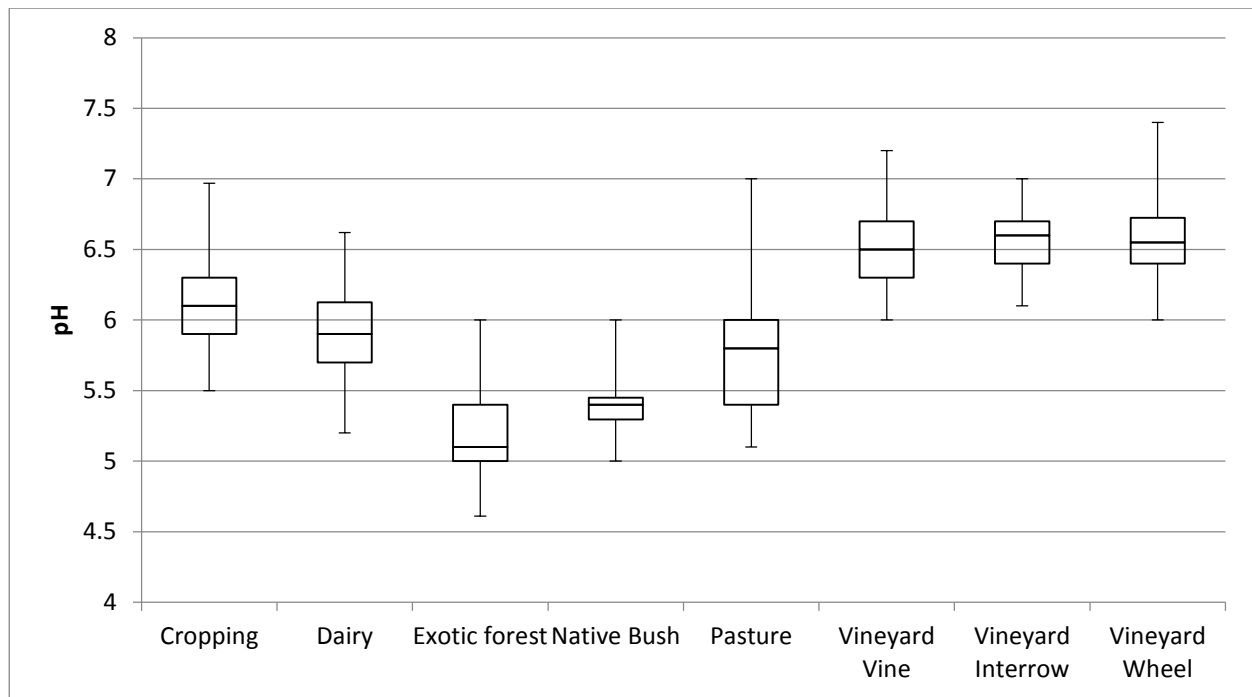


Figure 3: Soil pH by land use for all samples since 2000

3.2.2. Olsen P

Phosphorus is an essential nutrient for both plants and animals. Only a small amount of the total phosphorus in soil is in forms able to be taken up by plants (plant-available P). The Olsen P method is a chemical extractant that provides a reasonable estimate of the amount of plant-available phosphorus by measuring phosphate from soil solution and exchange surfaces (Olsen et al, 1954). Olsen-P can also provide an indicator for the risk of phosphorous loss to water. Phosphorus in run-off water is known to increase with increased Olsen-P values.

From the 40 Olsen P samples taken in 2017, concentrations varied from 3mg/L to 67 mg/L. The lowest values are found in forestry, dairy and pasture samples (Figure 4). Cropping was not included in figure 4 as there was only one sample point (31 mg/L). The highest values were found in vineyard samples. The maximum Olsen P target for all soils is set at 50 ml/L (Mackay *et al*, 2013). One vineyard site (3 samples) showed Olsen P values well in excess of this (58 to 67 ml/L).

Twenty-three sites had Olsen P values below the optimal for their land use. These sites may be operating at Olsen P values below concentrations considered optimal for maximum pasture/crop production. A number of these sites have low values because they are unimproved pasture (sites 14, 22 and 24). These sites have just never had fertiliser applied and their Olsen P value is consistently low over time.

Some sites are reflective of previous land use. Site 9 is a vineyard converted from dairying in the past year. The 2016 Soil quality report noted the high Olsen-P levels seen in dairy sites. Site 28 is similarly a recent vineyard conversion but from pasture, hence the low Olsen-P values.

Of the remaining low Olsen-P samples, some have lifted since their last sampling indicating the application of fertiliser (sites 2 and 27) and some have fallen probably as a result of removal of nutrients in products (site 4, 44). This leaves 11 sites with previously low values that have remained low at this sampling. Three of these sites are exotic forest and these values are expected to remain low. The final 7 samples are 6 vineyard and one dairy sample. These consistently low samples from sites (1, 4, 7 and 43) would suggest that fertiliser practice should be reviewed to ensure adequate productivity is being obtained from these sites.

The trends in the 2017 values are consistent with the longer-term samples (Figure 5). Farmed sites generally reflect higher Olsen P concentrations compared to unfarmed sites or sites with lower returns.

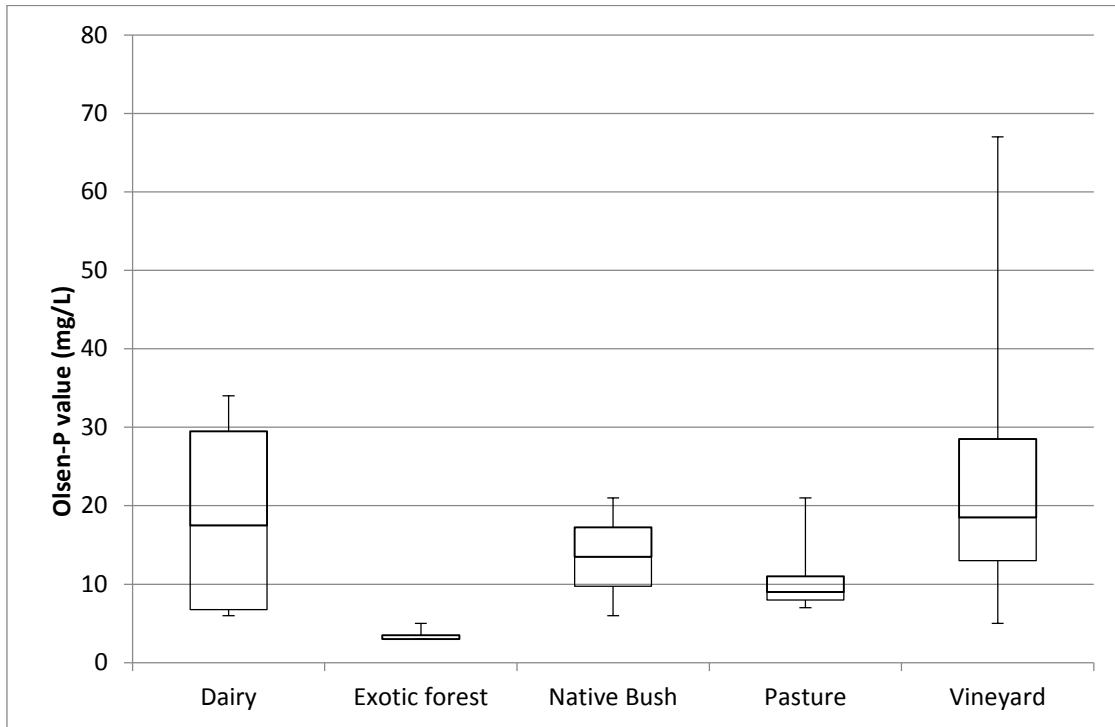


Figure 4: Olsen P values by land use for 2017 samples

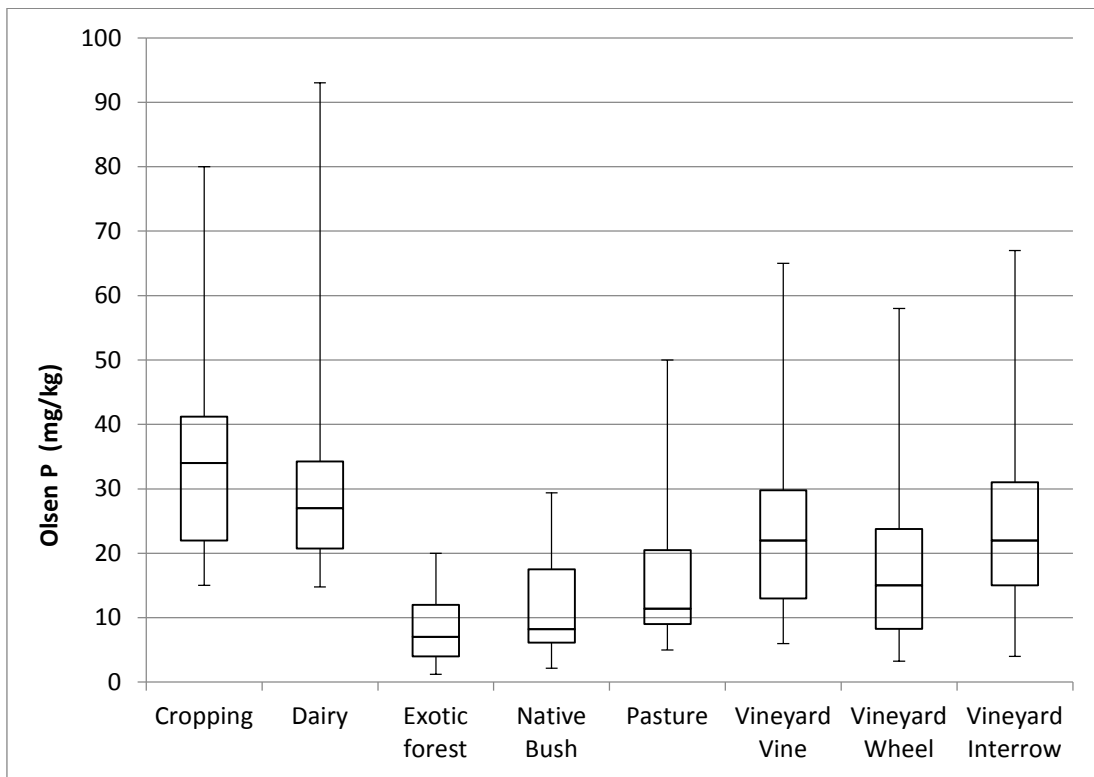


Figure 5: Soil Olsen P values for all samples since 2000

3.2.3. Trace Elements

Trace elements accumulate in soils either naturally through weathering of minerals contained in the soil parent material or from anthropogenic sources. While many trace elements are essential for healthy plant and animal growth, i.e. copper and zinc, at high concentrations in soils these can have a negative impact on soil fertility and plant and animal health. Furthermore, some trace elements, i.e. cadmium and arsenic are not essential in soils and their accumulation can also have a negative impact on soil, plant and animal health and in some cases, there is potential for them to accumulate in the human food chain.

Table 3 summarises trace element concentrations in soils from the monitoring sites. On average the 2017 sample concentrations were 3.95 mg kg^{-1} for arsenic, 0.13 mg kg^{-1} for cadmium, 19.1 mg kg^{-1} for chromium, 15.7 mg kg^{-1} for copper, 11.4 mg kg^{-1} for lead, 15.6 mg kg^{-1} for nickel, 0.12 mg kg^{-1} for mercury and 57.4 mg kg^{-1} for zinc. These **average** concentrations are within the suggested upper limits for trace elements in soils as suggested by the New Zealand Water and Waste Association (Appendix A). Concentrations are also similar to those that have been found in soils in other parts of New Zealand (Auckland Regional Council, 1999; Greater Wellington Regional Council, 2005; Canterbury Regional Council, 2006; Curran-Cournane & Taylor, 2012) and what has previously found in Marlborough (Gray C., 2011b).

For cadmium, average concentrations in farmed soils were approximately double typical background concentrations found in soils (0.2 mg/kg). Non-farmed soils such as native forest samples show only background levels of cadmium (Figure 6). The source of cadmium is most likely phosphate fertiliser which has been shown to contain cadmium as an incidental impurity (Longhurst, Roberts, & Waller, 2004).

The levels found in farmed soils are similar to those that have been found in the past for dairy pasture soils in Marlborough and what has been found elsewhere in New Zealand (Taylor, Kim, Hill, & Chapman, 2010). While no sites have exceeded the target values for cadmium (1 mg/kg) this year, one sample (Site 41) has reported a cadmium concentration above the suggested 0.6 mg kg^{-1} trigger value set in the Tiered Fertiliser Management System (TFMS) outlined in the New Zealand Cadmium Management Strategy (MAF, 2010). This dairy site has a history of high cadmium with its value falling from 1 mg/kg in 2008. The Olsen-P phosphate value from this site has risen during this time which would indicate use of fertiliser.

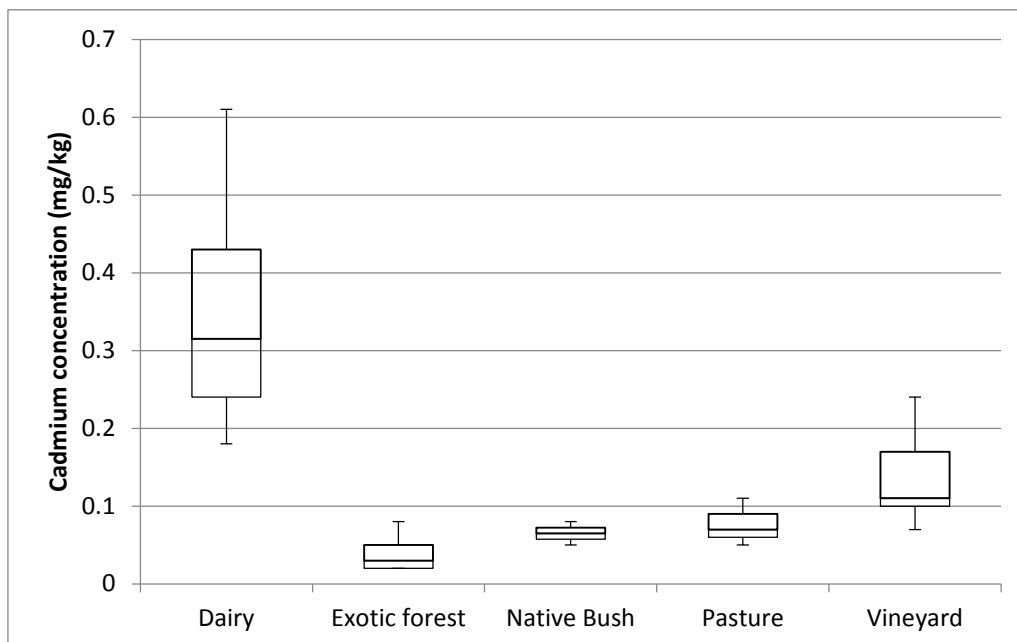


Figure 6: Soil cadmium concentrations by land use for 2017 samples

Overall, the 2017 results reflect the wider situation of cadmium concentration. Typically, farmed land uses have a higher cadmium concentration than non-farmed (i.e. forestry or native bush). Within the farmed land uses, the concentration of cadmium is generally higher in land uses that have higher value returns reflecting the frequency with which fertiliser is applied (Figure 7). While there is a wide spread of values, Dairy continues to have the highest cadmium concentrations indicative of that industry's historic reliance on phosphate fertilisers to boost pasture (and clover) growth.

The concentration of vineyard cadmium samples have lifted slightly this year with the conversion of site 9 to Vineyard. This site was a long-term dairy and had cadmium values of near 60 mg Cd/kg. Addition of this site has raised the maximum vineyard values but not changed the median or quartile values significantly.

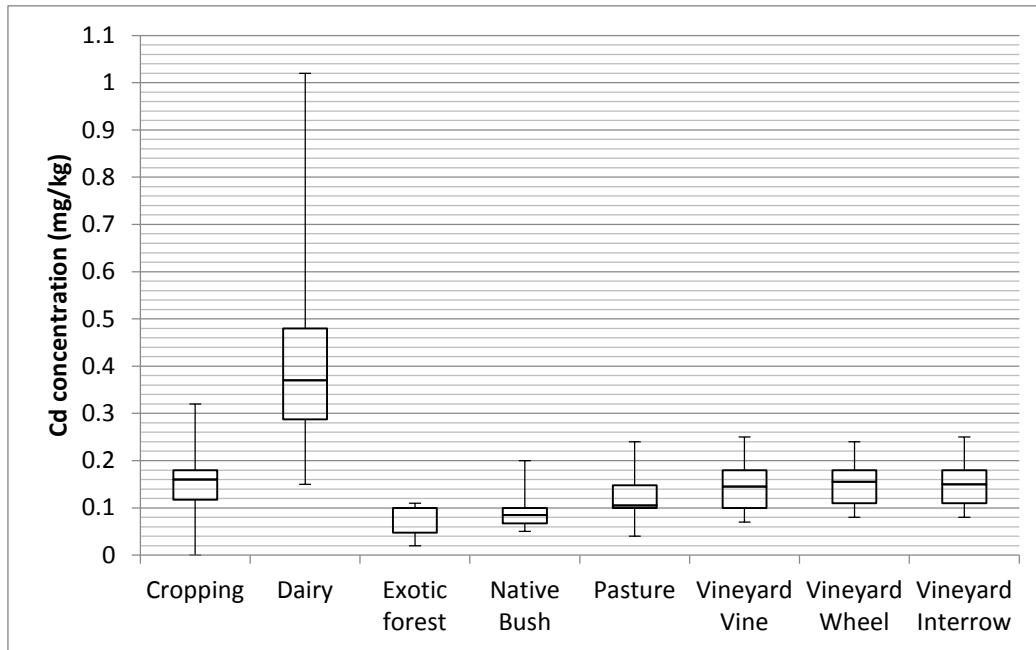


Figure 7: Soil cadmium concentrations by land use for all samples since 2000

Table 3 Soil chemical results. Soil pH, Olsen P and trace elements

Site	Soil type	Soil Order	Landuse	pH	Olsen P (mg/L)	Trace Elements							
						Zn (mg/kg)	Cu (mg/kg)	Cr (mg/kg)	As (mg/kg)	Pb (mg/kg)	Ni (mg/kg)	Hg (mg/kg)	Cd (mg/kg)
SOE_Soils_Site01a_vine	Wairau	Recent	Vineyard	6.3	23	68	21	22	5.8	11.9	26	< 0.10	0.13
SOE_Soils_Site01b_wheel	Wairau	Recent	Vineyard	6.5	13	59	20	20	4.4	12	25	< 0.12	0.11
SOE_Soils_Site01c_interow	Wairau	Recent	Vineyard	6.4	5	55	19	16.9	4	12.4	23	< 0.11	0.11
SOE_Soils_Site02a_vine	Wairau	Recent	Vineyard	6.4	20	64	17	18.9	4.3	12.4	25	< 0.10	0.2
SOE_Soils_Site02b_wheel	Wairau	Recent	Vineyard	6.7	28	63	16	19.5	4	12.1	26	< 0.11	0.22
SOE_Soils_Site02c_interow	Wairau	Recent	Vineyard	6.5	19	63	16	22	3.8	12.1	28	< 0.11	0.17
SOE_Soils_Site04a_vine	Renwick	Pallic	Vineyard	6.4	13	55	18	16	3.3	9.2	11.1	< 0.11	0.1
SOE_Soils_Site04b_wheel	Renwick	Pallic	Vineyard	6.5	19	57	21	11.5	3.5	8.9	8.5	< 0.11	0.09
SOE_Soils_Site04c_interow	Renwick	Pallic	Vineyard	6.3	18	54	20	11.6	3.5	9	7.8	< 0.11	0.09
SOE_Soils_Site05	Renwick	Recent	Cropping	6.5	31	67	12	15.6	3.5	11.2	10.7	< 0.11	0.13
SOE_Soils_Site06	Renwick	Pallic	Pasture	6	21	57	9	13.9	2.4	8.7	10.8	< 0.11	0.11
SOE_Soils_Site07a_vine	Paynter	Pallic	Vineyard	6.2	8	65	16	16.3	2.4	9.1	10.3	< 0.11	0.07
SOE_Soils_Site07b_wheel	Paynter	Pallic	Vineyard	6.4	11	59	11	13.4	2.2	8.3	8.9	< 0.10	0.08
SOE_Soils_Site07c_interow	Paynter	Pallic	Vineyard	6.3	7	63	14	15.5	2.7	9.4	10.8	< 0.10	0.08
SOE_Soils_Site08a_vine	Paynter	Gley	Vineyard	6.3	30	55	10	13.2	3.4	9.6	8.2	< 0.10	0.14
SOE_Soils_Site08b_wheel	Paynter	Gley	Vineyard	6.4	36	50	9	12.9	2.7	8.6	8.2	< 0.11	0.17
SOE_Soils_Site08c_interow	Paynter	Gley	Vineyard	6.1	21	48	9	13.8	3	9.5	8.9	< 0.10	0.15
SOE_Soils_Site09a_vine	Paynter	Pallic	Vineyard	6.2	65	79	20	24	5.3	15.9	15	< 0.11	0.21
SOE_Soils_Site09b_wheel	Paynter	Pallic	Vineyard	6.4	67	82	22	26	6	15.9	15.3	< 0.10	0.21
SOE_Soils_Site09c_interow	Paynter	Pallic	Vineyard	6.4	58	81	22	25	5.9	15.9	15.7	< 0.10	0.24
SOE_Soils_Site14	Seddon	Pallic	Pasture	5.9	8	64	11	23	3.7	10.1	15.6	< 0.10	0.09
SOE_Soils_Site17	Kaituna	Recent	Native Bush	5.4	21	62	13	19.3	3.1	12.9	10.5	< 0.11	0.08
SOE_Soils_Site19	Kenepuru	Brown	Native Bush	5.6	6	42	14	13.3	4.9	10.9	7.9	< 0.11	0.05
SOE_Soils_Site20	Kenepuru	Brown	Exotic Forest	5.4	3	26	9	10.3	2.1	7.2	5.7	< 0.11	0.04
SOE_Soils_Site21	Kenepuru	Brown	Exotic Forest	5.1	3	29	9	11.6	3.4	8.3	5.4	< 0.10	0.02
SOE_Soils_Site22	Kenepuru	Brown	Pasture	5.4	9	37	12	10.9	2.3	7.3	5.5	< 0.10	0.07
SOE_Soils_Site24	Wairau	Recent	Pasture	5.9	7	60	10	16.7	3.4	14.4	25	< 0.11	0.05
SOE_Soils_Site27a_vine	Motukarara	Gley	Vineyard	6.7	16	68	18	23	4.8	16	25	< 0.11	0.1
SOE_Soils_Site27b_wheel	Motukarara	Gley	Vineyard	6.7	30	65	18	21	4.8	16.8	22	< 0.10	0.1
SOE_Soils_Site27c_interow	Motukarara	Gley	Vineyard	7	16	65	19	22	4.5	15.9	24	< 0.12	0.12
SOE_Soils_Site28a_vine	Motukarara	Gley	Vineyard	6.2	15	59	15	23	5.1	15.3	27	< 0.10	0.11
SOE_Soils_Site28b_wheel	Motukarara	Gley	Vineyard	6.4	13	59	15	21	4.8	15.2	27	< 0.11	0.11
SOE_Soils_Site28c_interow	Motukarara	Gley	Vineyard	6.3	9	59	15	19.4	4.9	15.3	27	< 0.11	0.1
SOE_Soils_Site40	Kaituna	Brown	Dairy	5.7	34	60	14	18.4	3.6	11.9	10.6	< 0.11	0.37
SOE_Soils_Site41	Rai	Recent	Dairy	5.8	28	61	25	67	6.2	11.9	32	0.12	0.61
SOE_Soils_Site42	Pelorus Steepland	Brown	Exotic Forest	5.3	3	66	34	37	4.4	10.4	16.8	< 0.10	0.08
SOE_Soils_Site43	Pelorus Steepland	Brown	Dairy	5.4	6	61	20	24	4.8	11.7	10.6	< 0.10	0.18
SOE_Soils_Site44	Koromiko	Brown	Dairy	6.1	7	56	18	12.5	4.4	8.9	10.8	< 0.11	0.26
SOE_Soils_Site52	Tuamarina	Brown	Pasture	5.5	11	15	4	8.8	1.4	4.9	4.2	< 0.12	0.06
SOE_Soils_Site53	Tuamarina	Brown	Exotic Forest	4.8	5	38	13	13.7	5.5	9.8	10.6	< 0.10	0.02

Red cross indicates exceeds target range, red arrow below target range, yellow exclamation indicates exceeds TMFS level 1.

3.3. Soil Organic Matter Results

Results of soil organic matter analysis (anaerobically mineralisable nitrogen, total nitrogen, total carbon and C:N ratio) are reported in Table 2. Each of these organic matter properties is discussed individually. The target values appropriate to the relevant soil order can be found in Appendix A.

Recent work by Taylor, et al., (2017), Curtin, et al., (2017) and (Lawrence-Smith, McNally, Beare, & Lehto, 2018) has shown that hot water carbon and nitrogen extractions could provide a better soil quality indicator than the current set of organic matter indicators. In coming years, MDC will be adding hot water extractions to the analysis alongside the current measurements to help cross reference with existing data.

3.3.1. Anaerobically Mineralisable Nitrogen

Anaerobically mineralisable nitrogen (AMN) is a measure of the amount of nitrogen that can be supplied to plants through the decomposition of soil organic matter by soil microbes. It is a useful measure of soil quality that determines the ability of organic matter to store nitrogen. However, the amount of AMN has also been found to correspond with the amount of soil microbial biomass – hence it is also a useful indicator of microbial activity in soils (Myrold, 1987)

AMN can provide an indication of N loading in soil as organic matter and plant residues are mineralised (converted by microbes to mineral N). Mineralisation rates are strongly influenced by many factors such as temperature, moisture and C: N ratio. If the rate of mineralisation exceeds the rate of plant uptake, this will increase the amount of NO_3^- -N in soil solution (Havlin, Tisdale, Nelson, & Beaton, 2013). Increased soil solution N increases the risk of nitrate leaching. However, NO_3^- -N losses are also controlled by other factors such as soil texture and soil structure which affect the rate of water movement (drainage) in the soil and therefore the rate of NO_3^- -N loss. In addition, because soils are only sampled to the 10-cm depth, it isn't necessarily going to accurately reflect what happens to the NO_3^- -N further down the soil profile e.g. denitrification.

In 2017, anaerobically mineralisable nitrogen concentrations varied widely between sites with the lowest values found on unfarmed sites. Three sites had values which were higher than their target range and one was lower. In 2017, the dairy sites continued to have high readings due to the high rates of organic matter that accumulate in soils under dairy management (Figure 8). Typically, sites with higher inputs of organic matter such as pasture grasses, manure and urine have higher readings of AMN (Figure 9 – dairy, native bush, pasture). Given the higher AMN values on these sites, organic matter may be providing a large portion of soil solution N. Tools such as Overseer can be used to estimate the nitrogen flux in soil.

Sites with lower organic matter inputs or with a high level of soil disturbance will report lower levels of AMN. Increased soil disturbance increases oxidation of soil organic matter and this can be seen in both cropping and exotic forestry sites.

Few sites have fallen below the minimum AMN target level for their land use but clear differences are seen between land uses (Figure 9). Particularly striking is the lower AMN values within vineyards. Vineyard wheel tracks and inter-rows show similar AMN values but the area under the vines has noticeably lower AMN values. The continual use of herbicide in this area is probably limiting organic matter input. The long term effect of this will be to limit nitrogen availability in this area potentially leading to increased fertiliser use.

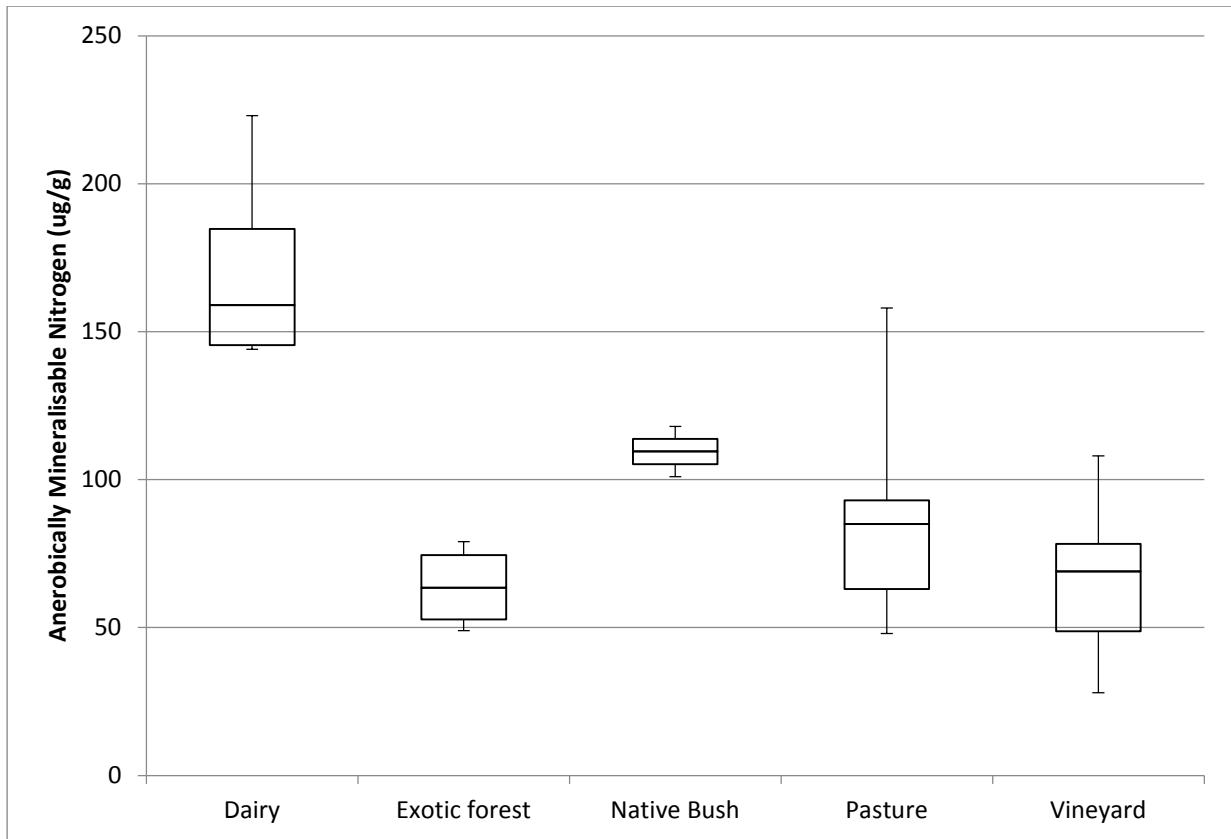


Figure 8: 2017 Anaerobically mineralisable nitrogen results

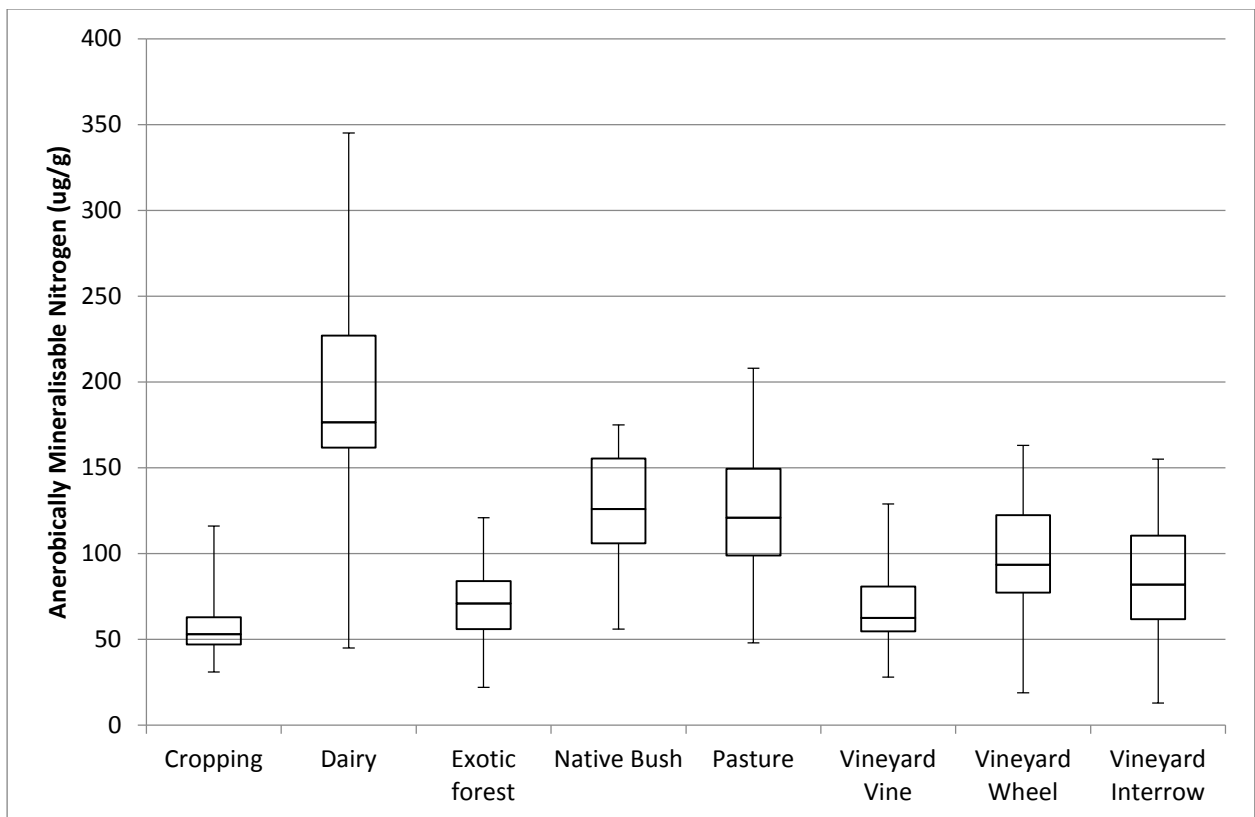


Figure 9: AMN concentrations by land uses for all samples since 2000

3.3.2. Total Carbon

Total carbon in soil includes carbonates and soil organic matter carbon. Typically, New Zealand soils contain only small amounts of carbonate; hence total carbon is generally considered a good measure of organic matter carbon in soil. Organic matter is important for soil health because it aids in the retention of moisture and nutrients, contributes to a stable soil structure, provides a source of energy for soil microbes and is a source of nutrients e.g. nitrogen, phosphorus and sulphur. In contrast, low soil C increases the risk of structural degradation in soils e.g. low aggregate stability, high bulk density, low air filled porosity, formation of surface crusts and compaction.

5 sites had total soil carbon contents below the acceptable target ranges for their respective land use activity. Of the sites that had total carbon below the target levels, four were vineyard sites and three were under-vine samples. It is normal for soils under long-term, high producing pastures to accumulate carbon. However, if organic matter inputs are low (as in the under-vine herbicide strip) or repeated cultivation occurs, total carbon levels will fall.

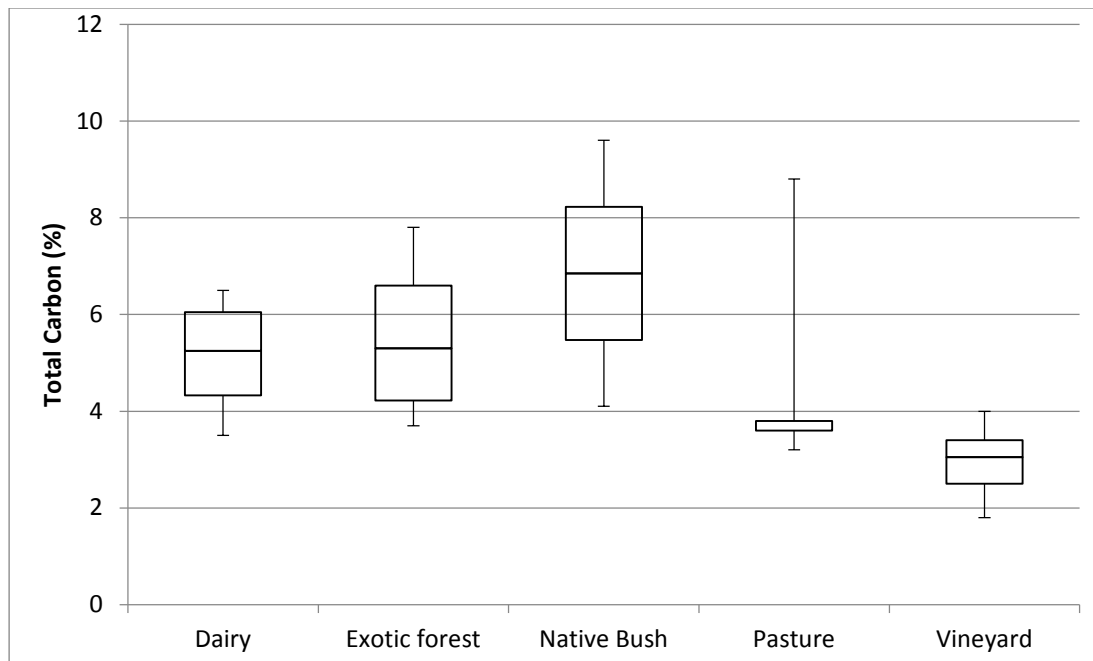


Figure 10: Total carbon values for 2017

It is clear from Figures 10 and 11 that organic matter accumulation and protection is greatest under native bush. This represents the carbon accumulation from deposition of organic matter over many thousands of years in some cases. The median figure for native bush of 5.6% could be taken as a guide to the pre-European level of soil carbon through much of lowland Marlborough.

The longer term figures (Figure 11) illustrate a similar situation to that of AMN. Land uses with high inputs of organic matter (dairy, forest, pasture) have higher levels of total carbon. Land uses that involve the disturbance of soil (cultivation) have reduced total carbon. It is interesting to note the difference between the exotic and native forest data (Figure 11). The exotic forest has lower total carbon compared to native bush most likely due to the intermittent soil disturbance that occurs during forest harvesting and establishment.

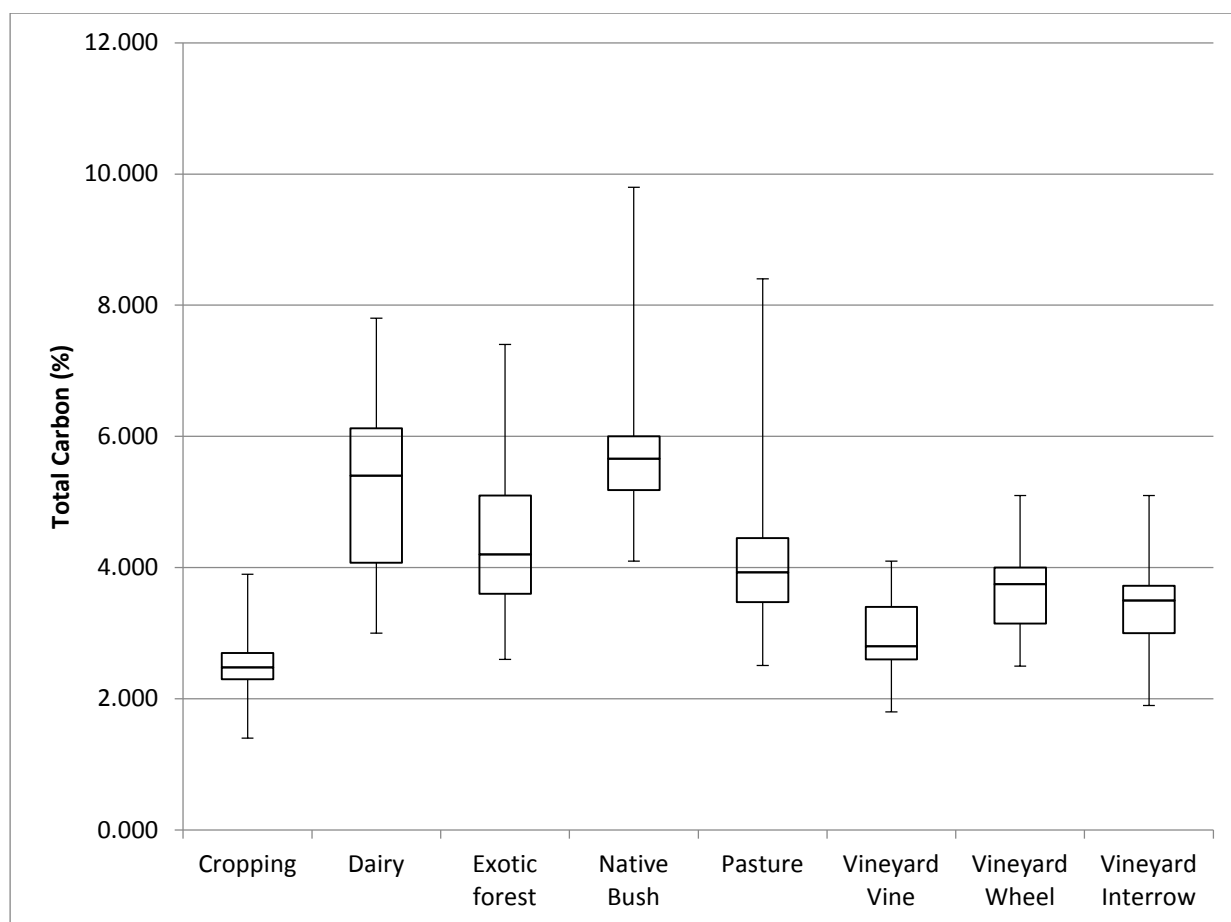


Figure 11: Total carbon by land use for all samples since 2000

3.3.3. Total Nitrogen

Nitrogen is an essential major nutrient for plants and animals, and the store of organic matter Nitrogen is an important measure of soil fertility. Typically, in topsoils, organic matter Nitrogen comprises more than 90% of the total Nitrogen. However, organic matter Nitrogen needs to be mineralised to inorganic forms (i.e. ammonium and nitrate) by soil microbes before it can be utilised by plants or lost from soil by leaching.

In 2017, all sites returned values above the target values for total Nitrogen. Note that no target values are set for total Nitrogen for cropping or horticulture (including vineyards). However, it is interesting to note the differences between the different vineyard samples. Inter-row samples would meet the pasture standard of 0.25% total N. The under-vine samples however would not meet this target and this is likely due to the low inputs of organic matter from the herbicide strip under the vines (Figure 12).

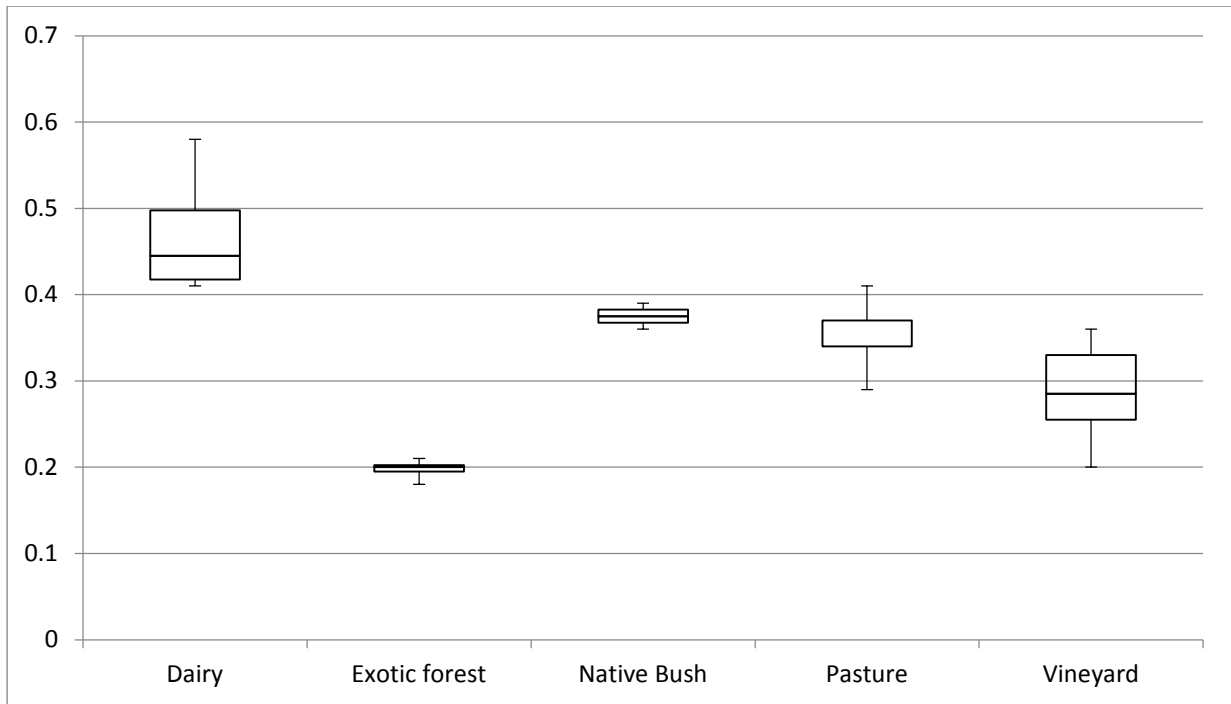


Figure 12: Total Nitrogen values for 2017

As total Nitrogen content is closely related to organic matter levels, soils with low inputs of organic matter or high loss rates caused by cultivation will have low total Nitrogen. This can be seen in Figure 13 in cropping, exotic forest and under-vine strips.

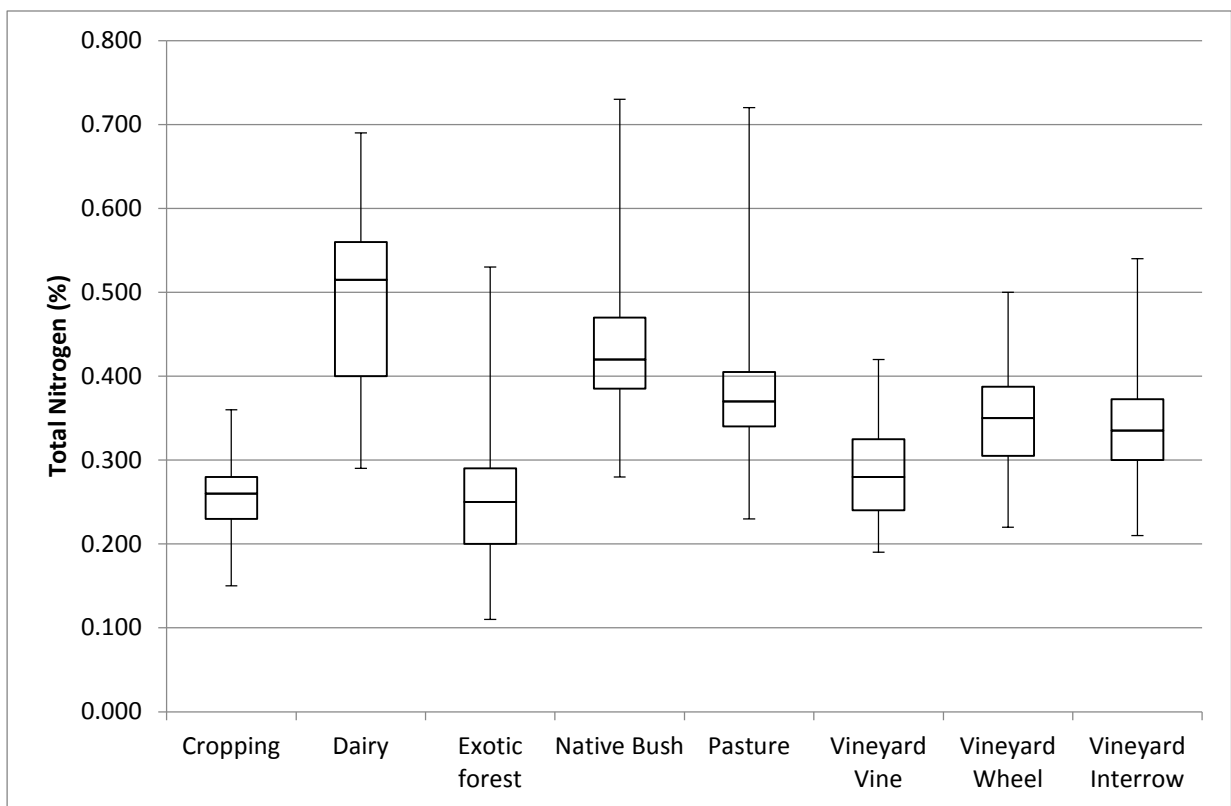


Figure 13: Total Nitrogen by land use for all samples since 2000

3.3.4. Carbon: Nitrogen Ratio

The balance of the amount of carbon to nitrogen in soil is called the carbon: nitrogen ratio (C:N). This ratio is important as a guide to the state of decomposition or likely ease of decomposition and mineralisation of nutrients i.e. production of nitrates and ammonium from organic residues in soils and is a measure of organic matter quality. It is therefore also a guide to the risk of N mobility (nitrate leaching) in soil.

Amongst the 2017 samples, 14 out of 40 samples had C:N ratios below 10:1 (Table 4) though most are close to 10:1. As C: N ratio increases above 10:1 (Nitrogen becomes scarce in relation to carbon), soluble Nitrogen is immobilised (taken up) by soil microbes, the soil solution N concentration falls and the risk of nitrogen leaching decreases (Havlin et al, 2013). Nitrogen cycling then becomes more dependent on microbial activity. Low C:N ratios (<10) may be of concern with regard to leaching of nitrate, as low ratios suggest the storage of N in organic matter may be reaching saturation. It has been estimated that within 40 years, most soils under intensive livestock farming would be near nitrogen saturation (Schipper, Percival, & Sparling, 2004). A nitrogen saturated soil can no longer store more organic nitrogen and potentially any additional nitrogen added will be lost from the soil and may ultimately accumulate in drainage waters and aquifers as nitrate. Hence monitoring the C:N over the medium to long term will provide useful information.

In comparison, native ecosystems with few nitrogen fixing plants and low nitrogen status, often have relatively high C:N ratios that have not received any additional nitrogen inputs (e.g. by stock grazing or fertiliser). Low nitrogen status is desirable for native ecosystems that have indigenous plants adapted to low nutrient conditions. Higher nutrient status may not be beneficial as this could encourage the growth of undesirable, weedy species. Exotic forest soils typically have C:N ratios >15 but are variable depending on whether they have been planted directly into cleared bush, a prepared site, second or third rotation, scrub, or former pasture. The current years data is in line with this with exotic forest reporting wide C:N ratios depending on the stage of forest development.

Table 4 Biological results, AMN, Total C, Total N and C: N ratio

Site	Soil type	Soil Order	Landuse	AMN µg/g	Total Carbon %	Total Nitrogen %	C:N Ratio
SOE_Soils_Site01a_vine	Wairau	Recent	Vineyard	72	2	0.21	9.7
SOE_Soils_Site01b_wheel	Wairau	Recent	Vineyard	70	3.2	0.33	9.8
SOE_Soils_Site01c_interow	Wairau	Recent	Vineyard	87	2.6	0.27	9.7
SOE_Soils_Site02a_vine	Wairau	Recent	Vineyard	53	1.9	0.2	9.7
SOE_Soils_Site02b_wheel	Wairau	Recent	Vineyard	61	1.9	0.21	9.1
SOE_Soils_Site02c_interow	Wairau	Recent	Vineyard	79	2.9	0.27	10.7
SOE_Soils_Site04a_vine	Renwick	Pallic	Vineyard	28	1.8	0.21	8.3
SOE_Soils_Site04b_wheel	Renwick	Pallic	Vineyard	43	2.5	0.26	9.8
SOE_Soils_Site04c_interow	Renwick	Pallic	Vineyard	45	2.5	0.26	9.5
SOE_Soils_Site05	Renwick	Recent	Cropping	52	2.4	0.24	10.3
SOE_Soils_Site06	Renwick	Pallic	Pasture	63	3.2	0.34	9.6
SOE_Soils_Site07a_vine	Paynter	Pallic	Vineyard	32	2.2	0.24	9.2
SOE_Soils_Site07b_wheel	Paynter	Pallic	Vineyard	54	3.1	0.33	9.4
SOE_Soils_Site07c_interow	Paynter	Pallic	Vineyard	72	3.7	0.36	10.2
SOE_Soils_Site08a_vine	Paynter	Gley	Vineyard	36	2.8	0.27	10.2
SOE_Soils_Site08b_wheel	Paynter	Gley	Vineyard	50	3.4	0.3	11.4
SOE_Soils_Site08c_interow	Paynter	Gley	Vineyard	45	4	0.36	11.2
SOE_Soils_Site09a_vine	Paynter	Pallic	Vineyard	60	3.4	0.3	11.3
SOE_Soils_Site09b_wheel	Paynter	Pallic	Vineyard	68	3.3	0.32	10.3
SOE_Soils_Site09c_interow	Paynter	Pallic	Vineyard	71	3.6	0.3	11.9
SOE_Soils_Site14	Seddon	Pallic	Pasture	93	3.6	0.34	10.4
SOE_Soils_Site17	Kaituna	Recent	Native Bush	118	4.1	0.36	11.4
SOE_Soils_Site19	Kenepuru	Brown	Native Bush	101	5.6	0.39	14.2
SOE_Soils_Site20	Kenepuru	Brown	Exotic Forest	73	4.4	0.2	22.3
SOE_Soils_Site21	Kenepuru	Brown	Exotic Forest	54	4.5	0.2	22.7
SOE_Soils_Site22	Kenepuru	Brown	Pasture	158	5.1	0.37	13.8
SOE_Soils_Site24	Wairau	Recent	Pasture	85	3.8	0.41	9.4
SOE_Soils_Site27a_vine	Motukarara	Gley	Vineyard	72	2.6	0.24	10.9
SOE_Soils_Site27b_wheel	Motukarara	Gley	Vineyard	78	3	0.31	9.8
SOE_Soils_Site27c_interow	Motukarara	Gley	Vineyard	88	3.4	0.33	10.3
SOE_Soils_Site28a_vine	Motukarara	Gley	Vineyard	92	3.1	0.27	11.7
SOE_Soils_Site28b_wheel	Motukarara	Gley	Vineyard	108	3.7	0.33	11.2
SOE_Soils_Site28c_interow	Motukarara	Gley	Vineyard	100	4	0.36	11
SOE_Soils_Site40	Kaituna	Brown	Dairy	172	4.6	0.42	10.8
SOE_Soils_Site41	Rai	Recent	Dairy	223	6.5	0.58	11.2
SOE_Soils_Site42	Pelorus Steepland	Brown	Exotic Forest	49	3.6	0.18	20.4
SOE_Soils_Site43	Pelorus Steepland	Brown	Dairy	144	5.9	0.47	12.5
SOE_Soils_Site44	Koromiko	Brown	Dairy	146	3.5	0.41	8.5
SOE_Soils_Site52	Tuamarina	Brown	Pasture	48	3.8	0.29	13.3
SOE_Soils_Site53	Tuamarina	Brown	Exotic Forest	79	3.7	0.21	17.2

AMN: Green tick= within range, red cross exceeds target range or red arrow under target range

Total C: Green tick= above minimum, red cross below minimum

Total N: Green tick= above minimum, No target values for horticulture or cropping

C:N ratio: Below 10 (green tick), reduced risk of leaching.

3.4. Soil Physical Results

3.4.1. Bulk Density

Bulk density is the weight of soil in a specified volume and provides a measure of how loose or compacted a soil is. Loose soils may be subject to increased risk of erosion, dry out quickly, and plant roots find it difficult to get purchase and absorb water and nutrients. In contrast, soils with a high bulk density are generally compacted and have poor aeration and are slow draining. The consequences of compacted soil may include reduced supply of air to plant roots, increased resistance to penetration that may limit root extension and germination, and reduced capacity of the soil to store water that is available to plants. Further, reduced water entry into the soil may increase water runoff over the soil surface (McLaren & Cameron, 1996).

Eight sites from the 40 samples from 2017 had bulk density values above the target ranges for the relevant soil orders (Table 5, Figure 14). Four of the eight are vineyard wheel track samples. All wheel track samples had higher bulk density values than the corresponding vine or inter-row samples. Only 3 wheel track samples were below the upper limit for their soil types and two of these were new vineyards (Site 9 and 28) recently converted from pasture. The remaining four out-of-range samples are the one cropping site (site 5) and 3 vine or inter-row samples (Sites 2 and 4). Both Site 2 & 4 are long-term vineyard sites. Interestingly, Site 1 is located close to Site 2 on the same soil type. However, Site one has bulk density readings within the target values. This would indicate that different vineyard management regimes can influence soil bulk density.

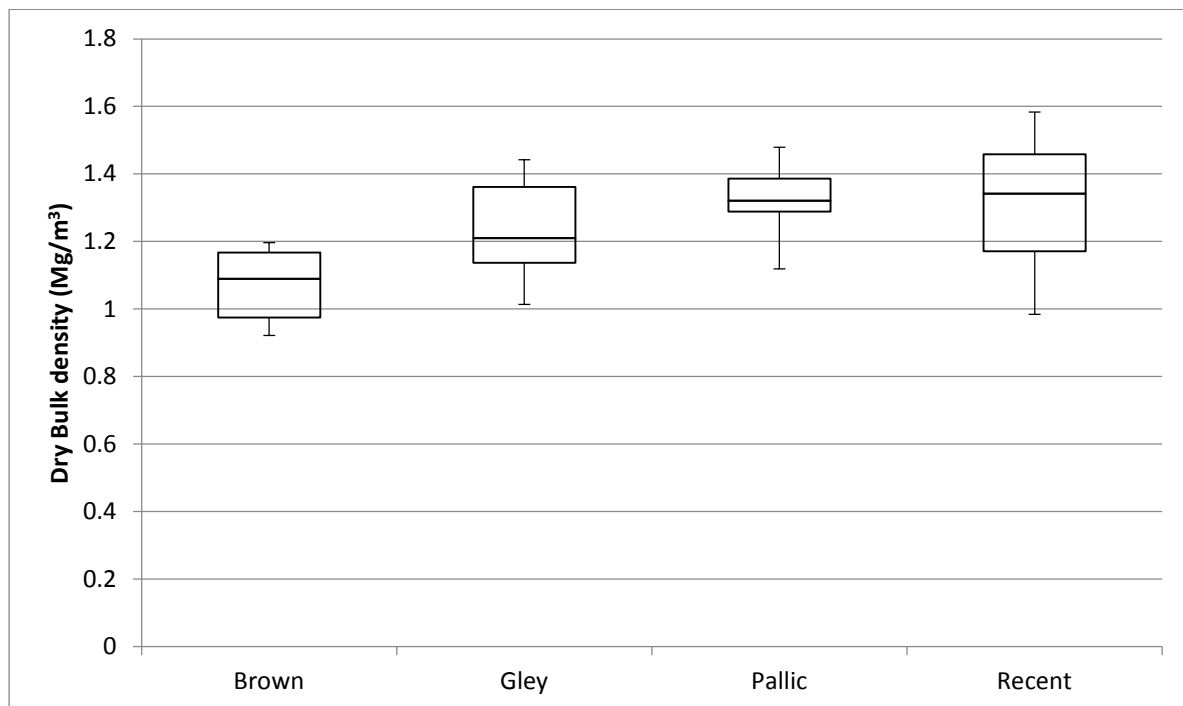


Figure 14: Dry bulk density by soil orders for 2017 samples

Figure 15 shows bulk density for different land uses since samples began in 2000. Bulk density values tend to reflect the level of farming activity. Intensive farming that involves soil disturbance, repeated trafficking by vehicles and livestock treading, all display higher bulk density readings. Low intensity sites show low bulk density readings with native bush again providing a baseline value. Dairy farms provide an interesting counterpoint here however. As will be seen in coming sections, the higher organic matter inputs into this system seems to protect the soil to some degree against developing higher bulk density despite the heavy treading caused by cattle. However, dairy soils are still regarded as compacted. This is because large pores are removed by the treading. Often this is insufficient to cause a lift in bulk density. The removal of these large pores contrasts with the regular vehicle trafficking seen in vineyards which remove all pore sizes leading to the increase in bulk density.

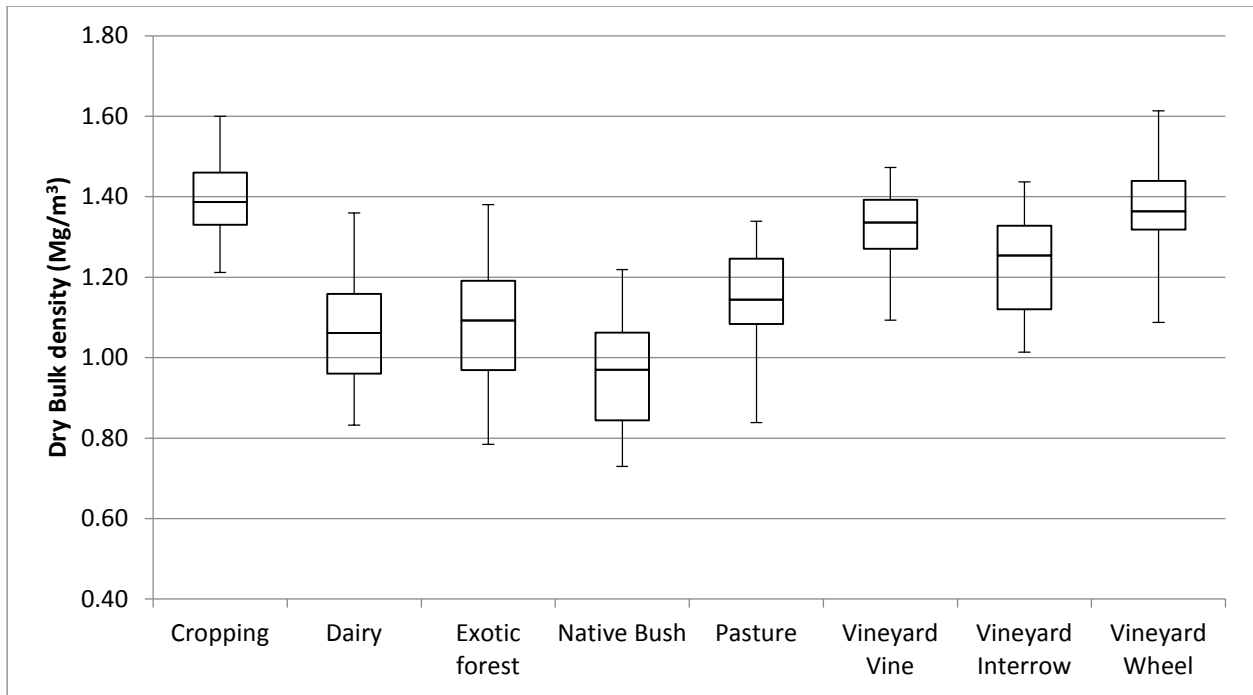


Figure 15: Soil dry bulk density values for all samples since 2000

3.4.2. Air Filled Porosity

Air filled porosity (AFP) is a measure of the proportion of large pores (macropores) in the soil. Macropores are important for penetration of air into soil, extension of roots down into the soil and drainage of water. Typically, macropores are the first to be lost when the soil is compacted. It is generally accepted that when air filled porosity represents less than 10% of the total soil porosity, plant growth will be affected (McClaren and Cameron, 1996)

Soil compaction has again been shown to be a major problem on Marlborough farms. 2017 results show dairy and vineyard wheel tracks are badly affected (Figure 16). Air filled porosity readings in past Soil Quality reports have identified compacted soils under all forms of farmed land in Marlborough. The 2017 samples show one of the clearest examples of soil compaction. Only forestry and pasture land uses show AFP readings consistently over the target limit. (Table 5). Low air filled porosity has been noted previously in Marlborough (Gray, 2011a) and has been observed in other regions of New Zealand (Taylor et al., 2010; Fraser and Stevenson, 2011; Stevenson, 2010; Sorensen, 2012). The Ministry of the Environment recently summarised nationwide regional council soil quality data and found only 22% of dairy sites met the required standard (MfE, 2016). The low values are likely related to heavy grazing or grazing under wet conditions where animal treading has effectively reduced the large pore fraction in soils.

A uniquely Marlborough problem is soil compaction under vineyard wheel tracks. This trend was discussed in the 2016 Soil Quality Report (MDC, 2017). Every 2017 vineyard wheel sample shows excessive compaction with one reporting a value of 0.9% (Site 2), an order of magnitude below the target limit of 10%. Compaction is also evident in other vineyard areas but seems to be dependent on the length of time a vineyard has been in production and the management regime. Older vineyards have reduced AFP in vine, wheel and inter-row samples (Sites 1, 2, 8) whereas younger vineyards often only show compaction under wheel tracks (Site 9, 28 4). Compaction is possibly increased where vineyard soils are stony due to the practice of using heavy rollers to bury stones.

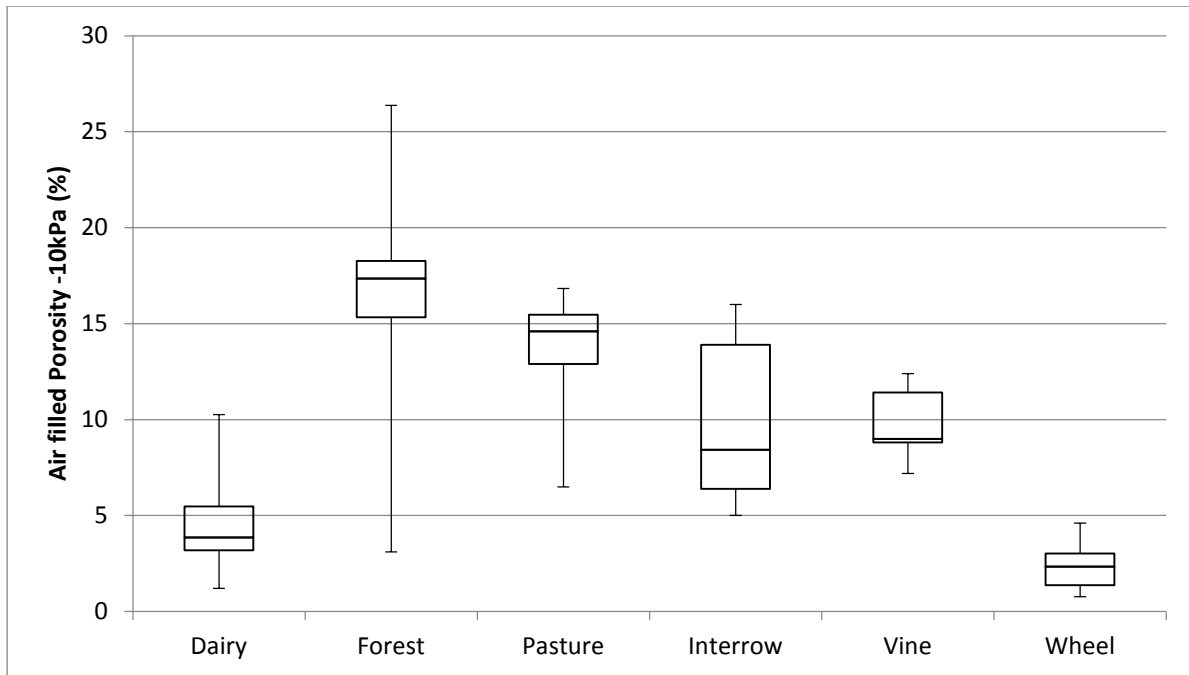


Figure 16: Air filled porosity by land use for 2017 values

Figure 17 shows air filled porosity data for all samples collected since 2000. While there is wide variance across the data, it is clear that three land uses have issues with compaction (cropping, dairy and vineyard wheel tracks). Interestingly, exotic forest regularly reports very high AFP readings. This may be a function of the irregular soil disturbance that occurs on these sites.

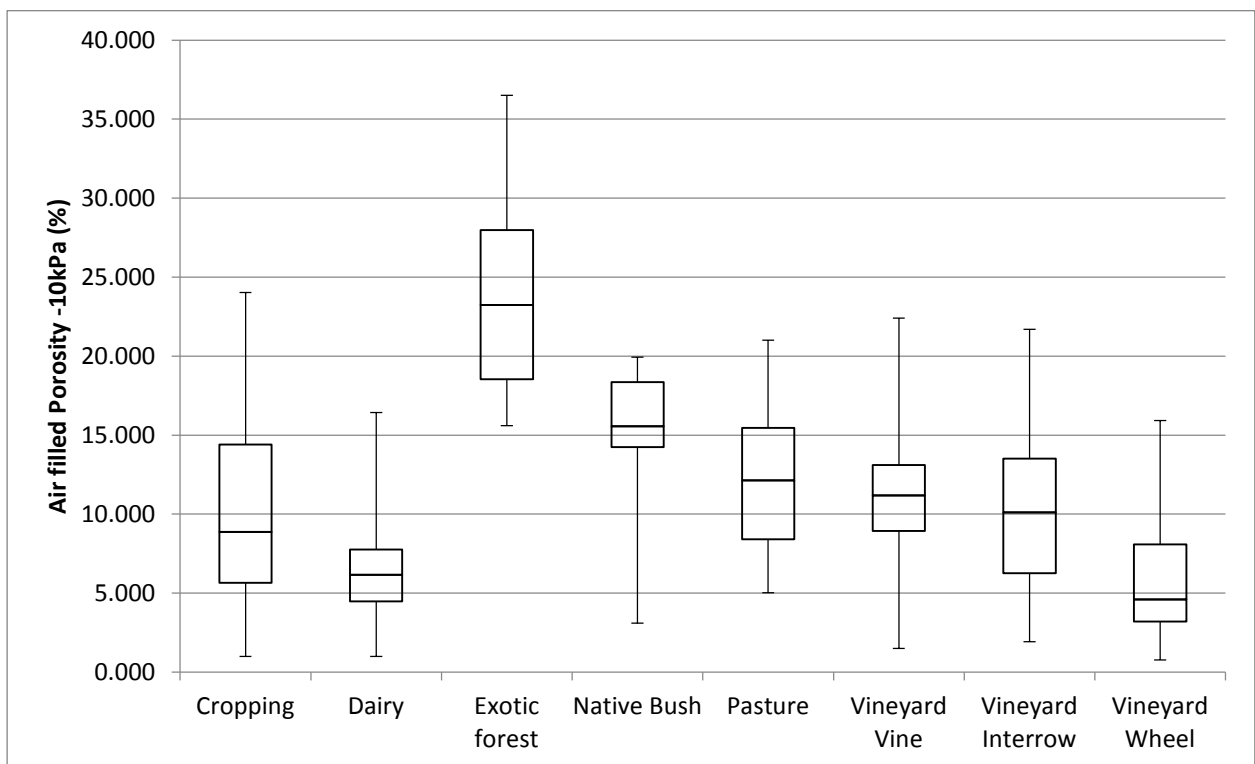


Figure 17: Air filled porosity by land use for all samples since 2000

3.4.3. Aggregate Stability

Aggregate stability refers to the ability of soil aggregates to resist disruption when forces such as rapid wetting and mechanical abrasion are applied. In general a soil with adequate amounts of soil organic matter will have stable soil aggregates and therefore a higher aggregate stability. A stable soil structure is important to allow water and air movement in soils and to minimise surface erosion (Mclaren & Cameron, 1996).

A single cropping site was sampled in 2017 and an aggregate stability measurement was taken (Table 5). Although there are no specific target ranges available for aggregate stability, generally any value below about 1.5 mean weight diameter (MWD) is considered low and likely to have a negative effect on crop production (Francis, Tabley, & White, 1991). With a value of 0.7 MWD, and combined with the low AFP and high bulk density, this site would be considered to be in poor condition and at risk of structural collapse. Remedial actions would include a pasture fallow period and addition of lime or gypsum to help rebuild soil structure.

Table 5 Physical characteristics of soils sampled in the Marlborough Region

Site	Soil type	Soil Order	Landuse	Dry Bulk Density (Mg/m3)	Macro Porosity (-5kPa) (% v/v)	Air Filled Porosity (-10kPa) (% v/v)	Aggregate Stability (MWD)
SOE_Soils_Site01a_vine	Wairau	Recent	Vineyard	✓ 1.37	7.37	✗ 8.97	
SOE_Soils_Site01b_wheel	Wairau	Recent	Vineyard	✓ 1.31	2.00	✗ 3.60	
SOE_Soils_Site01c_interow	Wairau	Recent	Vineyard	✓ 1.29	5.03	✗ 6.90	
SOE_Soils_Site02a_vine	Wairau	Recent	Vineyard	✗ 1.47	7.17	✗ 8.63	
SOE_Soils_Site02b_wheel	Wairau	Recent	Vineyard	✗ 1.58	0.10	✗ 0.90	
SOE_Soils_Site02c_interow	Wairau	Recent	Vineyard	✗ 1.43	3.50	✗ 5.00	
SOE_Soils_Site04a_vine	Renwick	Pallic	Vineyard	✗ 1.44	10.80	✓ 12.40	
SOE_Soils_Site04b_wheel	Renwick	Pallic	Vineyard	✗ 1.47	3.53	✗ 4.60	
SOE_Soils_Site04c_interow	Renwick	Pallic	Vineyard	✓ 1.32	11.37	✓ 13.43	
SOE_Soils_Site05	Renwick	Recent	Cropping	✗ 1.53	4.30	✗ 3.93	0.70
SOE_Soils_Site06	Renwick	Pallic	Pasture	✓ 1.33	14.30	✓ 16.83	
SOE_Soils_Site07a_vine	Paynter	Pallic	Vineyard	✓ 1.33	10.03	✓ 11.57	
SOE_Soils_Site07b_wheel	Paynter	Pallic	Vineyard	✗ 1.48	0.25	✗ 0.77	
SOE_Soils_Site07c_interow	Paynter	Pallic	Vineyard	✓ 1.30	6.35	✗ 6.20	
SOE_Soils_Site08a_vine	Paynter	Gley	Vineyard	✓ 1.37	5.60	✗ 7.20	
SOE_Soils_Site08b_wheel	Paynter	Gley	Vineyard	✗ 1.44	1.17	✗ 2.13	
SOE_Soils_Site08c_interow	Paynter	Gley	Vineyard	✓ 1.36	4.87	✗ 6.47	
SOE_Soils_Site09a_vine	Paynter	Pallic	Vineyard	✓ 1.20	7.67	✗ 8.87	
SOE_Soils_Site09b_wheel	Paynter	Pallic	Vineyard	✓ 1.29	0.40	✗ 1.53	
SOE_Soils_Site09c_interow	Paynter	Pallic	Vineyard	✓ 1.12	13.37	✓ 15.27	
SOE_Soils_Site14	Seddon	Pallic	Pasture	✓ 1.29	3.97	✗ 6.50	
SOE_Soils_Site17	Kaituna	Recent	Native Bush	✓ 1.13	2.40	✗ 3.10	
SOE_Soils_Site19	Kenepuru	Brown	Native Bush	✓ 0.99	12.13	✓ 14.67	
SOE_Soils_Site20	Kenepuru	Brown	Exotic Forest	✓ 1.09	16.80	✓ 18.57	
SOE_Soils_Site21	Kenepuru	Brown	Exotic Forest	✓ 1.19	15.57	✓ 17.30	
SOE_Soils_Site22	Kenepuru	Brown	Pasture	✓ 0.97	12.63	✓ 15.47	
SOE_Soils_Site24	Wairau	Recent	Pasture	✓ 1.05	11.77	✓ 14.60	
SOE_Soils_Site27a_vine	Motukarara	Gley	Vineyard	✓ 1.21	10.07	✓ 11.37	
SOE_Soils_Site27b_wheel	Motukarara	Gley	Vineyard	✓ 1.33	2.95	✗ 2.83	
SOE_Soils_Site27c_interow	Motukarara	Gley	Vineyard	✓ 1.14	8.63	✗ 9.97	
SOE_Soils_Site28a_vine	Motukarara	Gley	Vineyard	✓ 1.10	7.43	✗ 9.03	
SOE_Soils_Site28b_wheel	Motukarara	Gley	Vineyard	✓ 1.19	2.00	✗ 2.53	
SOE_Soils_Site28c_interow	Motukarara	Gley	Vineyard	✓ 1.01	14.00	✓ 16.00	
SOE_Soils_Site40	Kaituna	Brown	Dairy	✓ 1.17	0.10	✗ 1.20	
SOE_Soils_Site41	Rai	Recent	Dairy	✓ 0.98	2.17	✗ 3.87	
SOE_Soils_Site42	Pelorus Steepland	Brown	Exotic Forest	✓ 0.97	24.07	✓ 26.37	
SOE_Soils_Site43	Pelorus Steepland	Brown	Dairy	✓ 0.92	6.63	✓ 10.27	
SOE_Soils_Site44	Koromiko	Brown	Dairy	✓ 1.16	1.40	✗ 3.85	
SOE_Soils_Site52	Tuamarina	Brown	Pasture	✓ 1.09	9.43	✓ 12.90	
SOE_Soils_Site53	Tuamarina	Brown	Exotic Forest	✓ 1.20	15.00	✓ 17.40	

Bulk Density: Green tick- within range, red cross-exceeds range.

Air filled porosity: Green tick above target limit, red cross below target limit. 10% target limit for most land uses except for 8% in forestry.

4. Changes in Soil Quality through time

4.1. Introduction

The Soil Quality monitoring program seeks to fulfil the Marlborough District Council's legislative responsibilities under the RMA to report on the "life supporting capacity of soil" and to determine whether current practices will meet the "foreseeable needs of future generations". Soil quality and land use are also key drivers in water quality. As a result, it has been a long-term goal of the MDC to report on regional-scale changes in soil quality to inform debate about environmental impacts of human activities in our region.

To meet these goals and obligations, we seek to answer three questions related to indicators for soil health. These include:

- What is the state and change of soil quality (based on soil order or land use)?
- To what extent and timeframe will the level of indicator meet a target or critical level?
- What are the main drivers that influence state and change (anthropogenic and non-anthropogenic)?

Earlier Soil Quality Monitoring reports have not addressed changes in soil properties over time. Since the initial national 500 soils program was established in 2000, data has been gathered from 96 sites throughout Marlborough. With a 5-year re-visit interval between sampling, it has taken until 2016 for sufficient data to be gathered to allow some analysis of trends in soil quality.

The methodology for this process is to use 5 year rolling averages for each soil indicator on each land use (M. Taylor, Pers. Comm., 2017). All the data from the previous 5 years is included in any given years average (i.e. 2017 data is averaged with all data since 2013). This data is then presented by land use or soil order. The aim of this is to provide a regional overview of soil quality. This is a simple methodology and there are discrepancies in some data. These are noted where appropriate in the text. For some land uses (native & exotic forest especially) the number of samples and frequency of sampling is insufficient. Readers of the 2016 soil quality report will have noted the excessive influence of outlier values with some graphs unduly biased as a result. In this report, values for land uses with insufficient sample numbers (exotic and native forest) have been presented as a single point on the graph. This point is derived using all sample values since 2000 and is to provide a baseline value to compare with other land uses.

Since the recommendation in the 2016 report to review the number of sites and the frequency of monitoring in all land uses, the following changes have been made; no further sites have been established due to cost constraints and some sites have been deferred or brought forward to provide better temporal coverage for particular land uses. Ideally, the number of each type of site should be proportional to the regional land use percentage. Sampling frequency should reflect the nominal 5 yearly sampling cycles, that is, one-fifth of sites should be sampled per land use per year. It is Council's intention to submit the full data set to a statistician for analysis and redesign in the coming year (2018) to ensure the results are robust and different land uses and soil orders are adequately represented.

The three key issues identified in last year's initial long-term analysis are still relevant. These include the risk of nutrients being lost to waterways (especially Nitrogen and Phosphorous), the decline in soil organic matter under some land uses and the potential risks of trace element contamination for some land uses.

A final addition has been made to this year's report, changes in land use from pastoral to viticulture has become common place in Marlborough. Council wishes to understand the effect of these changes and the speed with which they occur.

4.2. Nutrient loss to water

Nutrients lost from land into waterways represent a detriment to both systems. Nutrients lost from land causes it to become less fertile and requires that fertiliser be used in order to maintain productivity. This becomes a significant expense to farmers. Often nutrients are manufactured and imported so require large amounts of energy to create and ship. When lost nutrients reach waterways, they can promote growth of unwanted biological growths including plants and bacterial slimes. These can choke waterways and cause loss of habitat for fish and other plant species. Loss of nutrients into groundwater can lead to human health issues when that water is used for drinking (Boyd, 2015) Given Marlborough's reliance of groundwater resources for both drinking and irrigation water this is a potentially serious issue for the region.

Nutrients are lost to water in two main ways. Leaching is the loss through soils beyond the reach of plant roots into deeper soil layers. These nutrients may eventually reach groundwater or drain into waterways. Total nitrogen and anaerobically mineralisable N is monitored to evaluate the risk leaching may pose to water. The second pathway of loss is via surface runoff. Phosphorous is most susceptible to this pathway as it is carried on soil particles. Assessment of soil compaction is also important to ascertain the ability of water to infiltrate or runoff any given soil surface. Bare or very loose soils are vulnerable to leaching and erosion (runoff). Compacted soils prevent water (and fertiliser) infiltration and promote runoff (Mclaren & Cameron, 1996)

4.2.1. Phosphorus risk

In General, soils in Marlborough have moderate P levels. Monitoring has shown that most sites have Olsen P levels well within the target ranges. Of note, are the elevated levels of Olsen P found in the more intensive farming systems of dairy, cropping and viticulture (Figure 5). These soils will pose more risk of runoff than the less intensive farming systems shown simply because of the elevated P concentration.

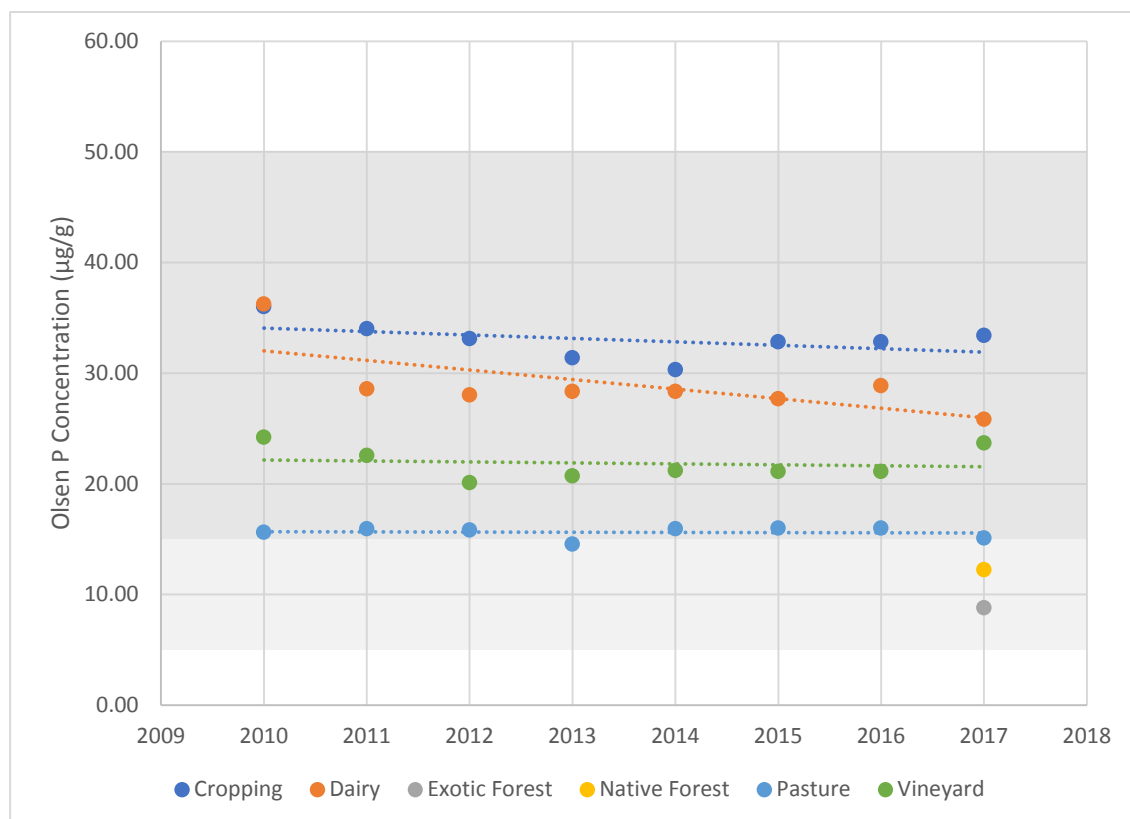


Figure 18: Regional Olsen P averages by land use

Cropping system risk depends mainly on the type of cultivation practice used to sow crops, the length of time land is left bare before sowing and weather during this time. Risk on dairy farms is posed by the

volume of dung left on the soil surface and the ability of the soil to assimilate this prior to rainfall or irrigation. Also of concern is the pugging of soils in wet conditions and residual grass cover left following grazing. These contribute to runoff risk of P by increasing soil compaction and by reducing the vegetation's ability to hold soil together under erosive conditions (Burgess, Chapman, Singleton, & Thom, 2000). Vineyard risk is lower but practices such as banding fertiliser, maintaining bare soil year-round (undervine, inter-row or both) and planting on slopes can increase P runoff risk.

There has been extensive national and international research to show that as soil P concentrations increase, the risk to waterways can also increase (McDowell, Drewry, Carey, Paton, & Condrón, 2003). On the back of these findings, a range of P mitigation strategies have been identified and tested to minimise P loss from soil to water. Some of these include achieving the optimal soil P test, use of low solubility P fertilisers, sediment traps, grass buffer strips, constructed wetlands, and application of amendments to sorb P in soil and drainage water (McDowell & Nash, 2012). Regular soil testing, and implementation of nutrient budget and management plans will help minimise excessive nutrient accumulation in soils and potential losses from soils and this is advocated to land managers.

The long-term trend in phosphate is generally stable for most land uses (Figure 18). A slight downward trend can be seen for dairy farms but this is within the margin of error for the data. Combined with the soil compaction data discussed earlier, this means phosphate loss risk is moderate but stable.

4.2.2. Nitrogen risk

The risk nitrogen poses to water quality is assessed by two tests. The total N test reports the complete content of N in the soil. This includes both the mineral and organic matter content. Anaerobically mineralisable N reports the ability of soil microbes to make soluble N by decomposing organic matter in the soil.

We see in Figures 19 & 20 that farm systems that involve animals (dairy and pasture) report higher rates of AMN and total N compared to non-animal farm systems (cropping, viticulture, forestry). This reflects increased fertiliser input, the increased production of easily decomposed organic matter (dung) and mineral N in urine. While both production systems are well within the target ranges on a regional basis, these measures can be highly variable on a spatial (farm to farm, paddock to paddock) and temporal (day to day, season to season) basis (Havlin et al, 2013). Elevated levels in these farm systems indicate that they pose greater risk to water quality than the non-animal systems. When variables such as slope, seasonal weather conditions, stocking rate, effluent disposal regimes, fertiliser application rates and frequency are included, there are likely be locations that do exceed the target ranges.

Non-animal farm systems (cropping, viticulture and forestry), show total N and AMN levels toward the bottom of the target bands. As will be seen in section 4.3, this is a result of lower organic matter content in these soils. It should be noted that cropping and horticulture have no general target ranges specified for total N. This is due to the large number of possible crops, each with its own target range. The lower levels of AMN found in the non-animal systems is likely to reduce the soils ability to produce nitrogen from organic matter. To compensate, farmers will likely require increased nitrogen fertiliser inputs. This may lead to increased risks to water from nitrogen loss depending on management practices such as application rates and timing.

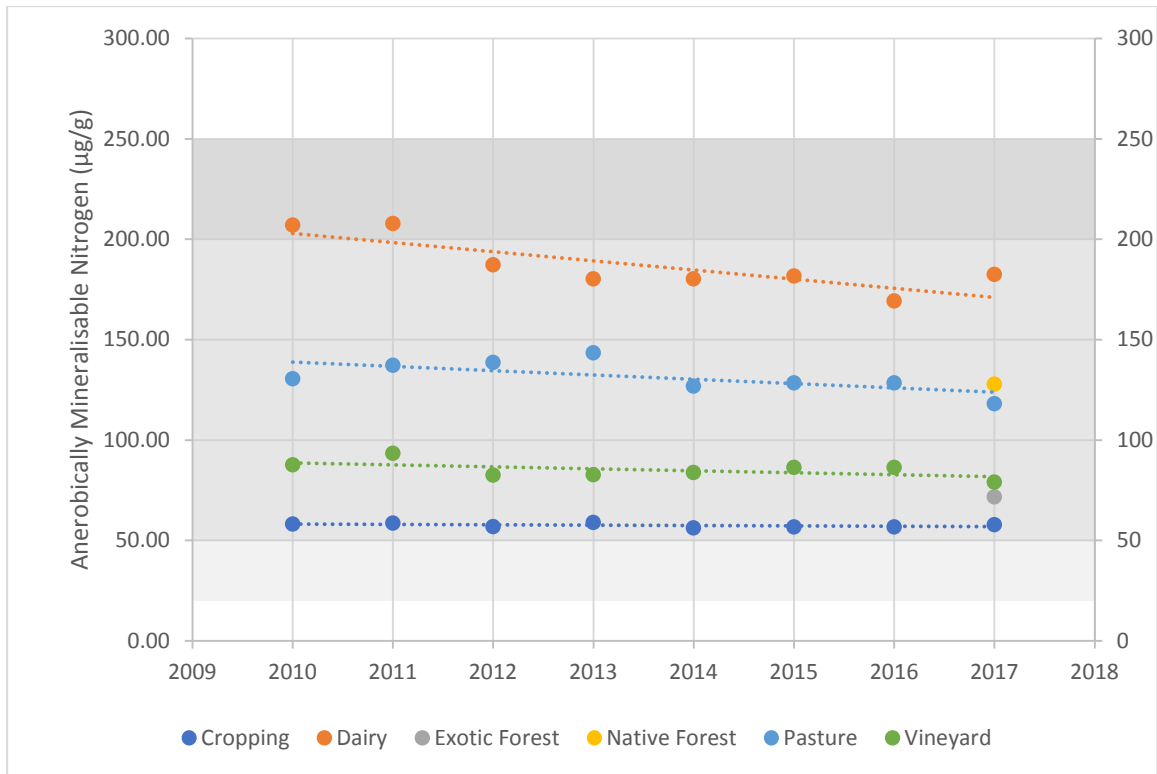


Figure 19: Anaerobically mineralisable nitrogen by land use

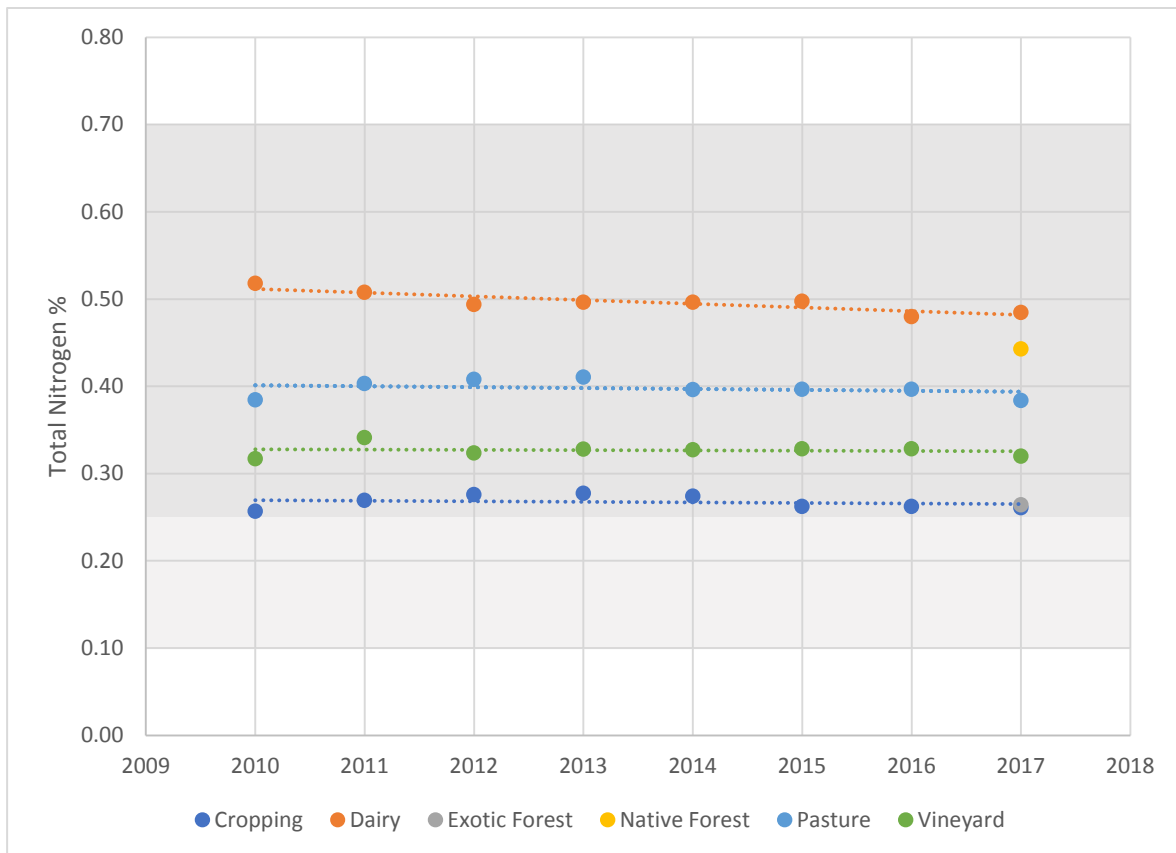


Figure 20: Total nitrogen by land use

4.2.3. Soil compaction risks

Soil compaction increases the risk of nutrient loss to water due to its role in increasing runoff. Soil compaction is measured by bulk Density and air filled porosity. Bulk Density measures the weight of a given volume of soil. It includes the pore space in that volume of soil and is strongly influenced by management practices that compact the soil (reduce pore space). Air filled porosity measures how much of the soil is normally filled by air (as opposed to water) at field capacity and represents a pore size of approximately 30µm in diameter (McLaren & Cameron, 1996).

There are a range of potential soil, plant and environmental effects of soil compaction/pugging. One of the most important is the effect on crop/pasture production. For example, animal grazing and treading, particularly in wet conditions, can affect pasture yield directly through leaf burial in mud, crushing, bruising and a reduction in dry matter production (Nie, Ward, & Michael, 2001). For both crops and pasture, indirect effects include; restriction of root penetration and radial growth of roots, reduced aeration, increased water logging potential due to slower drainage, reduced nutrient availability and water infiltration leading to reduced water storage in a soil. Reduced infiltration of water increases the potential for surface runoff of water. This runoff contributes increased risk of flooding. If runoff contains nutrients i.e. N, P or contaminants (i.e. bacteria), this may negatively impact on stream and lake water quality (Nguyen, Shealth, Smith, & Copper, 1998; McDowell et al, 2003).

The long-term trends in soil compaction in Marlborough mirror national trends (MfE 2016. pp. 84). Farmed systems have higher bulk density and lower air filled porosity (AFP) compared to non-farmed (forest) systems. Figures 15 and 17 illustrate these differences. Cropping and viticulture report the most compact soils but for different reasons. Cropping soils have the highest bulk density readings but very low AFP (with large variability in samples). This would indicate that both large and small pore spaces have been damaged by repeated cultivation. Cropping soils are also vulnerable to soil erosion when soils are cultivated prior to planting.

Soil compaction in viticulture is driven by trafficking of wheel tracks along rows (Figures 23 & 24). This repeated trafficking has removed the large soil pores but not small soil pores hence the lower bulk density readings compared to cropping soils (Figure 21 and 22). It should be noted that Soil Quality measurements are confined to in-vineyard sites. Vineyard headlands could reasonably be expected to have similarly compact soils due to high vehicle traffic. This would increase the area vulnerable to runoff. It is noted that vineyard wheel tracks and cropping seem to be showing improvement in AFP readings over time. This may reflect changes in practice around 2010 in viticulture to maintain grass coverage of wheel tracks and later in cropping. However for cropping, this trend may be a relic of inadequate sample size.

Both dairy and pasture systems show reasonable bulk density readings but very low and declining AFP (Figure 21 & 22). This will be due to treading damage by livestock compacting the large soil pores but not small pores. Combined with the raised levels of nitrogen and phosphate noted above, Marlborough's soil compaction presents a quite high risk of nutrient loss to water. Even though both N & P are within the target ranges, the level of soil compaction increases the risk of loss of these nutrients to water. Risk is increased when other factors such as slope, seasonal weather conditions, stocking rate, effluent disposal regimes, fertiliser application rates and frequency are considered.

There are a number of potential mitigation options that can be employed to prevent or minimise the effects of soil compaction. For pasture soils, some practices could include on/off grazing of animals; grazing wetter paddocks before the wet part of the season; maintaining good pasture cover which gives better protection against pugging; installing drainage in some areas; use of feeding platforms and/or standoff areas; decreasing winter stock numbers and moving stock onto well drained soil types off-site (Burgess et al, 2000). For cropping soils, maintaining practices that increase soil organic matter are important as well as minimising activity on soils during wet soil conditions that will compress and disrupt soil structure (Ghani, Mackay, Clothier, Curtin, & Sparling, 2009). For viticulture, mitigation is more difficult due to the need to drive rows frequently for various canopy management operations. Maintaining grassed wheel tracks and using mechanical loosening techniques may help in the short-term. Longer term solutions include raising soil organic matter and calcium levels and changing management techniques to minimise trafficking (multifunction machinery, over-row machinery).

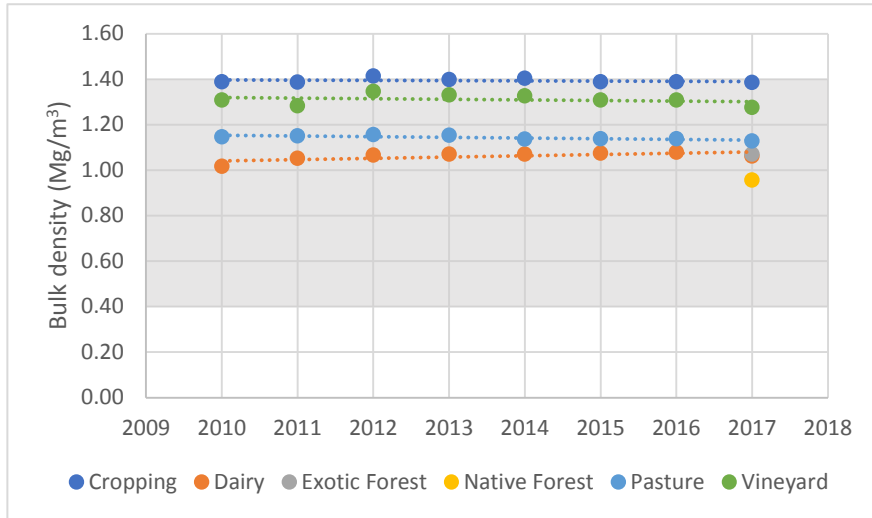


Figure 21: Change in bulk density for all land uses

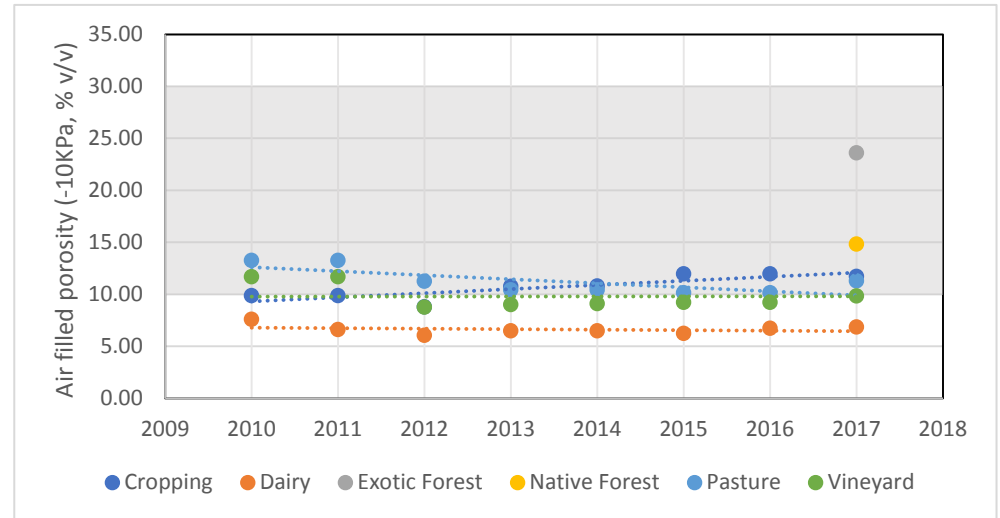


Figure 22: Change in AFP for all land uses

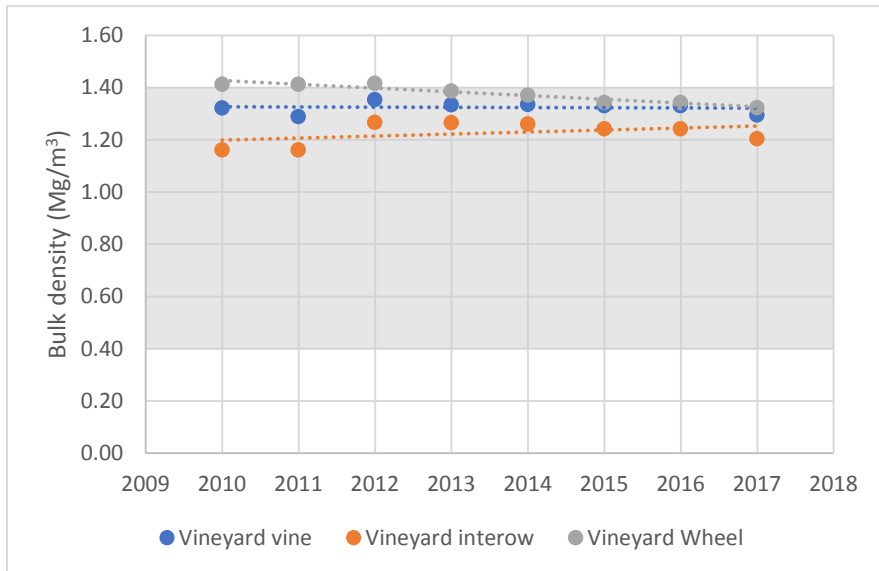


Figure 23: Change in vineyard bulk density

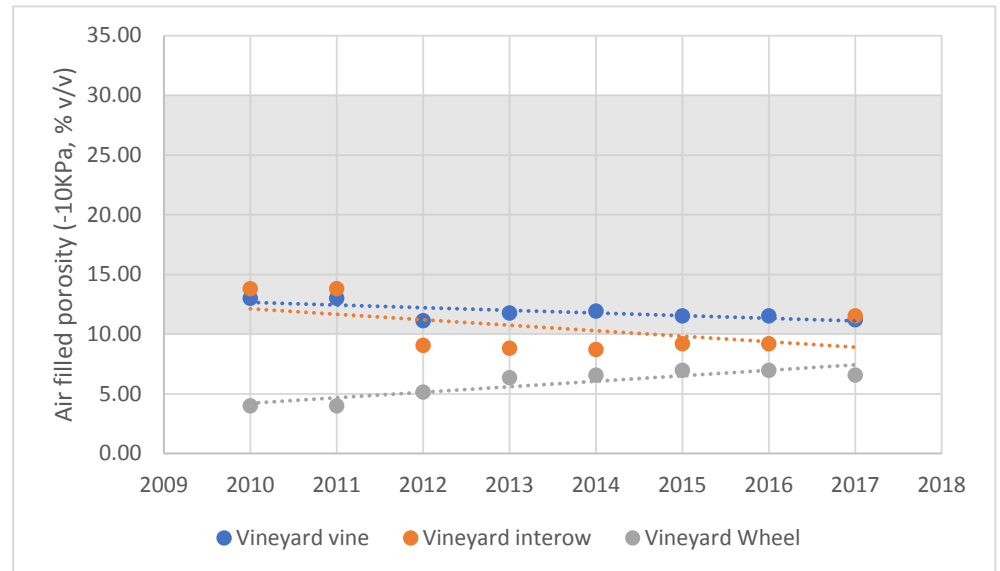


Figure 24: Change in vineyard AFP

4.3. Loss of Soil Organic Matter

Soil organic matter plays a significant role in the structural stability of soils as well as provision of nitrogen and carbon for use by soil microbes and plants. Low soil carbon (organic matter) increases the risk of soil structural degradation in soils e.g. low aggregate stability, high bulk density, low AFP, formation of surface crusts (Plate 2). In turn, poor soil structure can negatively affect things like soil aeration, drainage, water infiltration rates, water holding capacity, seed germination etc. In addition, loss of soil organic matter reduces the soils ability to retain nutrients from leaching and hold soil particles against runoff or erosion (Ghani, et al, 2009). These changes all have implications both for farm productivity and water quality.



Plate 2. Compacted topsoil at one of the cropping sites sampled with low soil carbon content (2012). Note the surface crust which reduces water infiltration, can increase surface run-off and reduce seed germination.

The indicator for organic matter status is total carbon. While this indicator has not dropped below the target values for any land use, and is showing little change over time, it is noticeable that farmed land uses have lower organic matter levels than native forest (Figure 25). We could regard the higher native forest level of around 6% as the pre-farming benchmark for soil organic matter. It is interesting to note the difference between total carbon content of exotic and native forest soils. Exotic forest reports carbon levels around 60% of native forest levels. This is most likely due to historic land clearance and erosion prior to exotic forest planting.

One land use (cropping) reports consistently low organic matter levels and this may have serious implications for soil and water quality. Cropping sites have the lowest carbon contents of the measured land uses. These results are consistent with trends observed during soil quality monitoring studies in both the Waikato and Wellington regions (Taylor, 2015; Sorensen, 2012) where cropping sites had depleted soil carbon contents compared to carbon at native vegetation sites. Most of the cropping sites had soil carbon contents at the lower boundary of their target range. Land managers need to adopt cultural practices that increase the amount of soil carbon, either by increasing carbon inputs or reducing the rate of decomposition of carbon. Such practices include residue management practices that maximise carbon returns to the soil, grow cover crops rather than leaving land bare over winter, include a pasture phase in rotations or adopt minimal tillage (Francis et al., 1991). These practices all help to reduce leaching and runoff and as such have a beneficial effect on downstream water quality and soil organic matter levels.

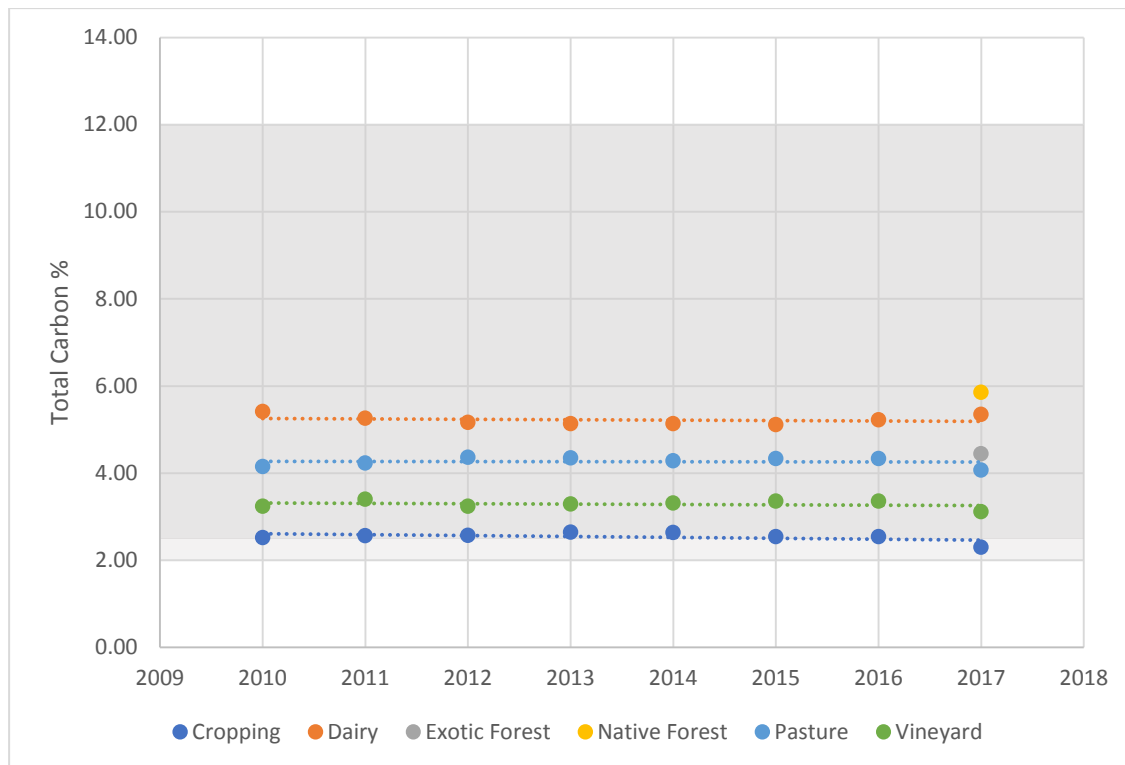


Figure 25: Total carbon by land use

In comparison to cropping sites, the dairy sites had higher total carbon content. It is well understood that soil under pasture will accumulate carbon. If the pasture is under a higher rainfall regime, irrigated, and fertilised, production of organic matter is increased and rates of carbon accumulation increase in response. This carbon can replace that lost through cultivation, decomposition, respiration and consumption.

It is the intention of the Council to introduce a hot water carbon test into the Soil Quality Monitoring program in the coming years. This test can help to determine the quality of the carbon in soils as well as the quantity. This will provide more information to help guide land management decisions.

4.4. Trace Element Contamination

The Soil Quality Monitoring program reports on many different trace elements found in soils (Gray, 2007a, Gray, 2007b). Many of these elements are toxic elements that are known to cause human and animal health problems (e.g. lead, mercury, cadmium and arsenic). The purpose of this is to inform Council of the risks of contamination from these elements. Monitoring has shown that there is little trace element contamination evident in most Marlborough soils.

The only other trace element of concern is cadmium. As a contaminant of phosphate fertiliser cadmium accumulates through time and is of concern for future land use change. A number of land uses (viticulture, cropping, pasture) show a slow increase in their cadmium content over time (Figure 26). The Tiered Fertiliser Management Strategy (TFMS) is a system for managing soil cadmium concentrations with different types of management action. For soils with cadmium concentrations up to 0.6 mg kg^{-1} (Tier 1) there are no limits on phosphate fertiliser application, but there is a recommendation that soils are tested for cadmium every five years. For soils which exceed 0.6 mg kg^{-1} but are below 1 mg kg^{-1} (Tier 2), phosphate fertiliser application rates are restricted to a specific set of products and application rates to manage cadmium accumulation to ensure cadmium concentrations don't exceed acceptable thresholds within the next 50 years. For soils which exceed 1 mg kg^{-1} but are below 1.4 mg kg^{-1} (Tier 3), application rates are further managed by use of a cadmium balance program to ensure that cadmium does not exceed an acceptable threshold within 50 years. While the monitoring of soil cadmium is the responsibility of regional councils, the implementation of these strategies is the responsibility of the fertiliser industry.

At current rates, the TFMS strategy Tier one level (0.6 mg kg^{-1}) would not be exceeded by viticulture, cropping or pasture land uses before 2067 (using the 50-year threshold time). However, because different land uses have different Maximum Residue Levels for cadmium, land use change could lead to contamination. For example, a soil that has accumulated cadmium under a pasture or vineyard regime that is then converted to vegetable production (cropping) may have sufficient cadmium to cause contamination problems in product. Understanding this, and given the high levels seen in some sites, it is suggested that land users test their soils for cadmium regularly and prior to land use change.

The situation with dairy cadmium levels is more problematic. The regional average levels are already concerning. See section 3.2.3. It should be noted that while the dairy trendline is down at present, there is considerable statistical error in this (see Figure 26). Only minor changes in future sample results could cause the trendline to shift up or down. If dairy cadmium levels were to increase at the same rate as Pasture levels are increasing, TFMS tier one would be exceeded around 2030.

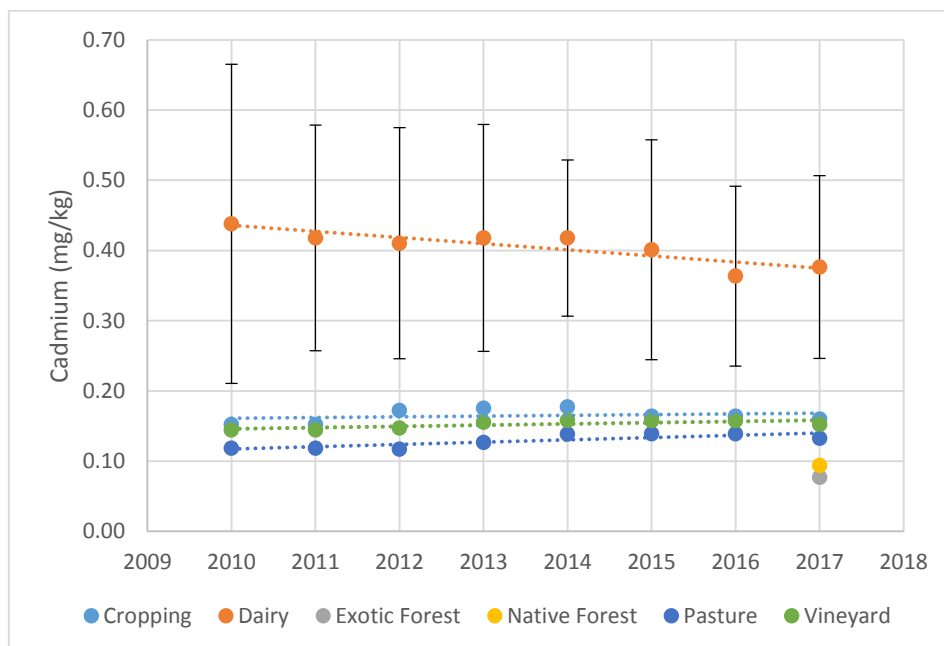


Figure 26: Cadmium levels by land use

4.5. Land use changes

The growth of the viticulture industry has seen large areas of former pastoral and orchard land converted to grape production. A number of changes in soil quality were expected as a result of these changes. These likely include increases in Olsen P, and bulk density alongside, declines in organic matter and AFP.

In 2017 two sites (Site 9 and 28), were identified that had recently converted to viticulture. Both are long standing Soil Quality Monitoring sites. These sites were sampled using the normal vineyard protocol. The results for site 28 are compared to a paired site that has been in viticulture since 2007 (Site 27). Site 9 is compared to Site 3 which is an ungrazed pasture site on the same farm and soil type.

This data should be viewed with caution. The data set is small. Each data point is based on only 3 replicates per year leading to a considerable margin of error. Note that measurements of wheel ruts only began in 2012. There are considerable time gaps between collection of individual data points. In future, as more land use change sites are identified, the data from these will be amalgamated and presented together in order to reduce error.

4.5.1. Land use change – Site 28

Site 28 was formerly grazed pasture. The soil type is Motukarara silt loam. It is paired with an adjacent vineyard (Site 27) on the same soil. The two sites are approximately 300 metres apart. Site 28 was converted to viticulture in 2016. During this time it was fully cultivated a number of times. When taking samples, the soils were noticeably softer and easier to work compared to older plantings. The site had around 18 months for the plantings to become established prior to the soil quality samples being taken in 2017.

While a number of soil test values have changed during this process, changes in bulk density, air filled porosity and total carbon stand out. Figure 27 shows bulk density for Site 28 starts low (0.96 Mg/m^3) as measured in 2008 consistent with a long-term pasture situation. This drops to 0.92 Mg/m^3 by 2013. This situation most likely persists until the cultivation of the pasture in 2015. After this time the bulk density has risen (although a drop and then steep rise is more likely) and the three samples begin to separate. Inter-row values stay relatively low, under-vine values rise with wheel tracks becoming rapidly compacted. A similar pattern has occurred for Site 27 with the three samples also separating out with increasing density of inter-row, vine and wheel samples.

Air filled porosity for these sites shows related behaviour. As bulk density rises, the soil becomes more compact and there is less pore space in the soil. In Figure 28 we can see that Site 27 and 28 have both commenced with high AFP values. The site 27 value is possibly the result of cultivation of the soil following the establishment of the vineyard but Site 28 will have high AFP due to its long-term pasture. In both cases the AFP has fallen following the establishment of vineyard. If Site 27 is a reliable guide, it would seem that under-vine and inter-row areas while declining may settle at around 10 % AFP while wheel track values will decline to below 5 %AFP.

This would indicate that compaction occurs over the whole vineyard area following establishment and is in line with the bulk density trends. We see a separation of the three samples into very compact wheel tracks and vine/inter-row areas with lower, albeit acceptable, AFP levels. This mirrors trends seen in bulk density.

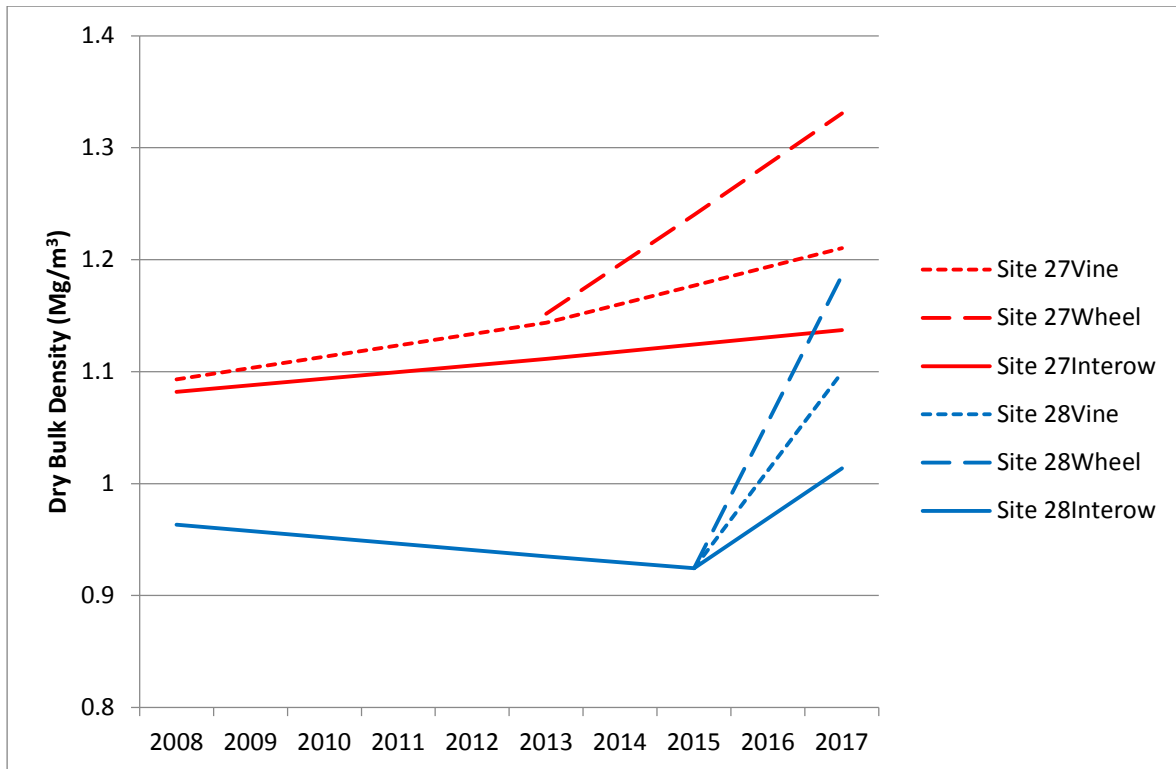


Figure 27: Bulk density of Sites 27 and 28 following vineyard establishment

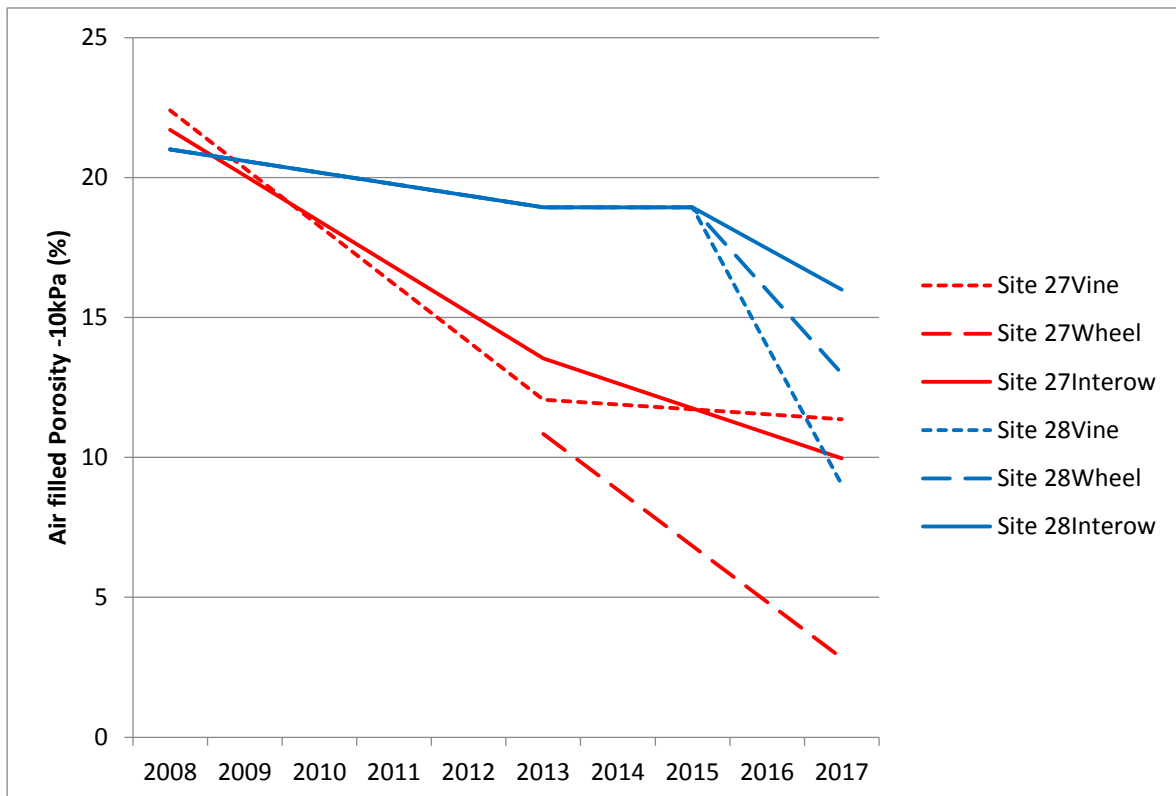


Figure 28: Air filled porosity for Sites 27 and 28 following vineyard establishment

The changes to the physical aspects of the soil following the establishment of vineyard would seem logical given the practices employed in this work. The repeated cultivation is used to reduce weed competition, improve vine root penetration and allow establishment of a permanent sward. Of interest in

this is the effect on soil organic matter. Theoretically, it should be possible to measure reductions in total carbon during the establishment phase and a slow rebuilding of this as the vineyard and sward establish themselves. Figure 29 shows the results of total carbon analysis. It is clear that vineyard establishment reduced total carbon on both sites and no recovery is yet evident on the older Site 27. Interestingly, the split between the three sample locations is evident again with under-vine areas having the lowest total carbon. Inter-row samples show the highest carbon levels with Wheel between these two values. Wheel tracks may have increased amounts of protected carbon due to the compaction. Compacted soils have slow oxygen penetration and this slows the oxidation of carbon.

The changes noted on sites 27 and 28 underline the results shown elsewhere in this report. Under-vine carbon levels are low, soils are very compact (especially under wheel tracks) and soil bulk density is high. We can now see that these may be a result of establishment practice and low organic matter inputs. It is also likely that due to the loss of organic matter, increased and continual compaction and, with low organic matter inputs, that soil structure may take some time to recover to former levels. It is likely that some parts of vineyards will never recover while vines are in place.

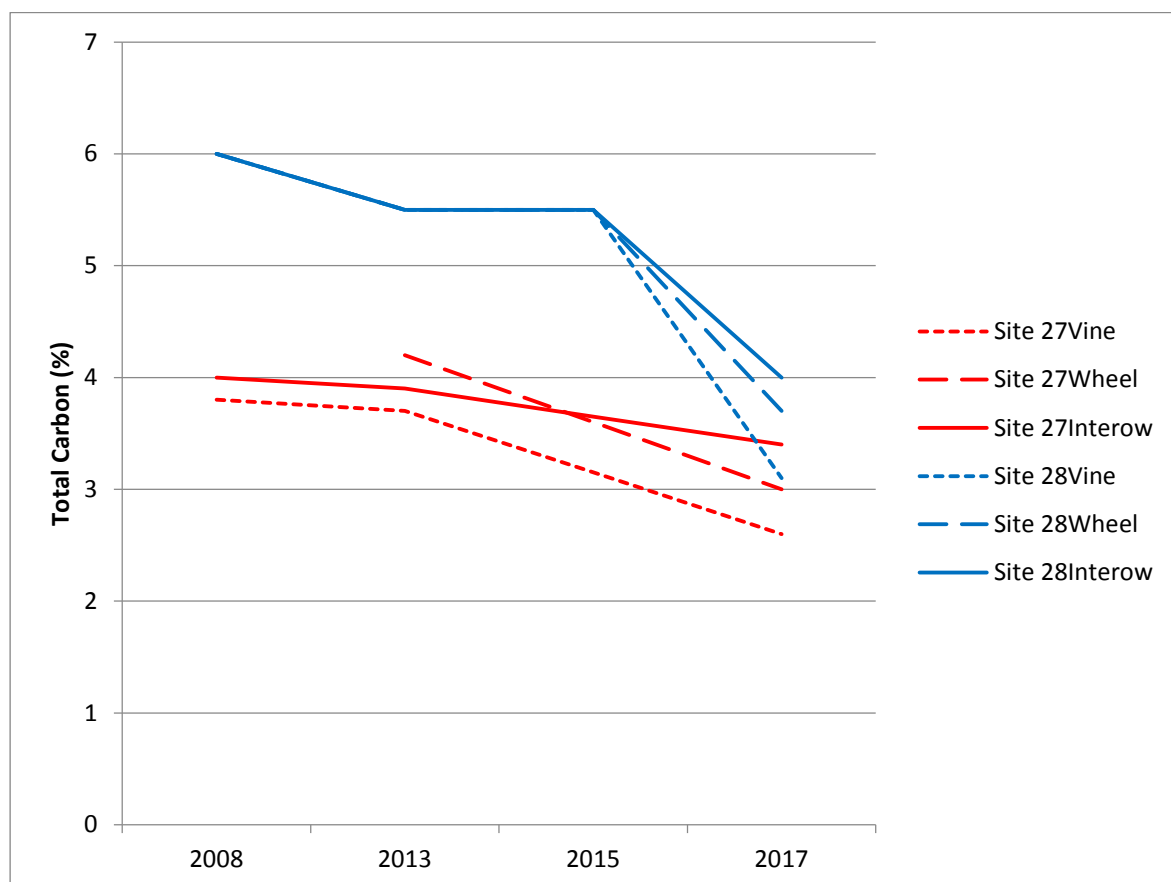


Figure 29: Total carbon for Sites 27 and 28 following vineyard establishment

A number of mitigation techniques could help to address this situation. Following establishment, managers could engage in actively lifting soil organic matter levels. This could include the use of compost under vines or allowing weeds to grow over the herbicide strip in winter. As an example, to lift the 2017 Site 28 vine measurement (3.1%) back to the 2015 value (5.5%), addition of 46.8 tonnes of carbon per hectare would be required. This would equate to incorporating around 140 tonnes of compost per hectare.

4.5.2. Land use change – Site 9

Site 9 has been farmed as a dairy farm for many years but was converted to viticulture in 2016. The site is located on Paynter soils described as a Typic Orthic Gley soil. This soil is deep clay with poor drainage. There is no matching pair for this soil so it is compared with the average values for dairy over the entire study.

Figure 30 shows the air filled porosity following conversion to viticulture. Site 9 starts at a value consistent with the average AFP for dairy. A measurement in 2012 reported a value of <1. This can be explained by either flaws in the sampling (likely) or analysis (unlikely) or that the soil had been badly pugged immediately prior to sample collection. This will mean that the 2012 value is probably non-representative (it lies well outside the margin for error for that years samples).

Following conversion in 2016, the site then reported values similar to those found at Sites 27 & 28 and consistent with the wider group of vineyard measurements for AFP. Inter-row areas were loose following repeated cultivation, under-vine areas were around the target value of 10 % and wheel tracks were badly compacted. For this level of compaction to occur so soon after establishment must be of concern to the grower as this will certainly be limiting root penetration for the establishing vines. Given this is a poorly drained site, root depth will be quite shallow due to high winter water tables further restricting root volume. This could lead to summer water stress and root rots. The loose soil found in the inter-row is also a potential erosion concern.

As seen in Figure 30, the inter-row AFP matches that of native forest. While this may seem to be a good comparison, the native forest has a total carbon level that is 2.2% higher than that for the Site 9 inter-row. What this implies is the AFP in the native sites is created from natural processes such as earthworm burrows and by high organic matter inputs. As such it will be resistant to erosion and compaction. In contrast, the high AFP in Site 9 inter-row is probably due to mechanical soil loosening (cultivation). Most likely the value will fall in coming years as the soil re-compacts. However, in the meantime, this loose soil will be more vulnerable to erosion.

Other test values for this site show similar trends to those seen in Site 27 and 28. This data is not presented.

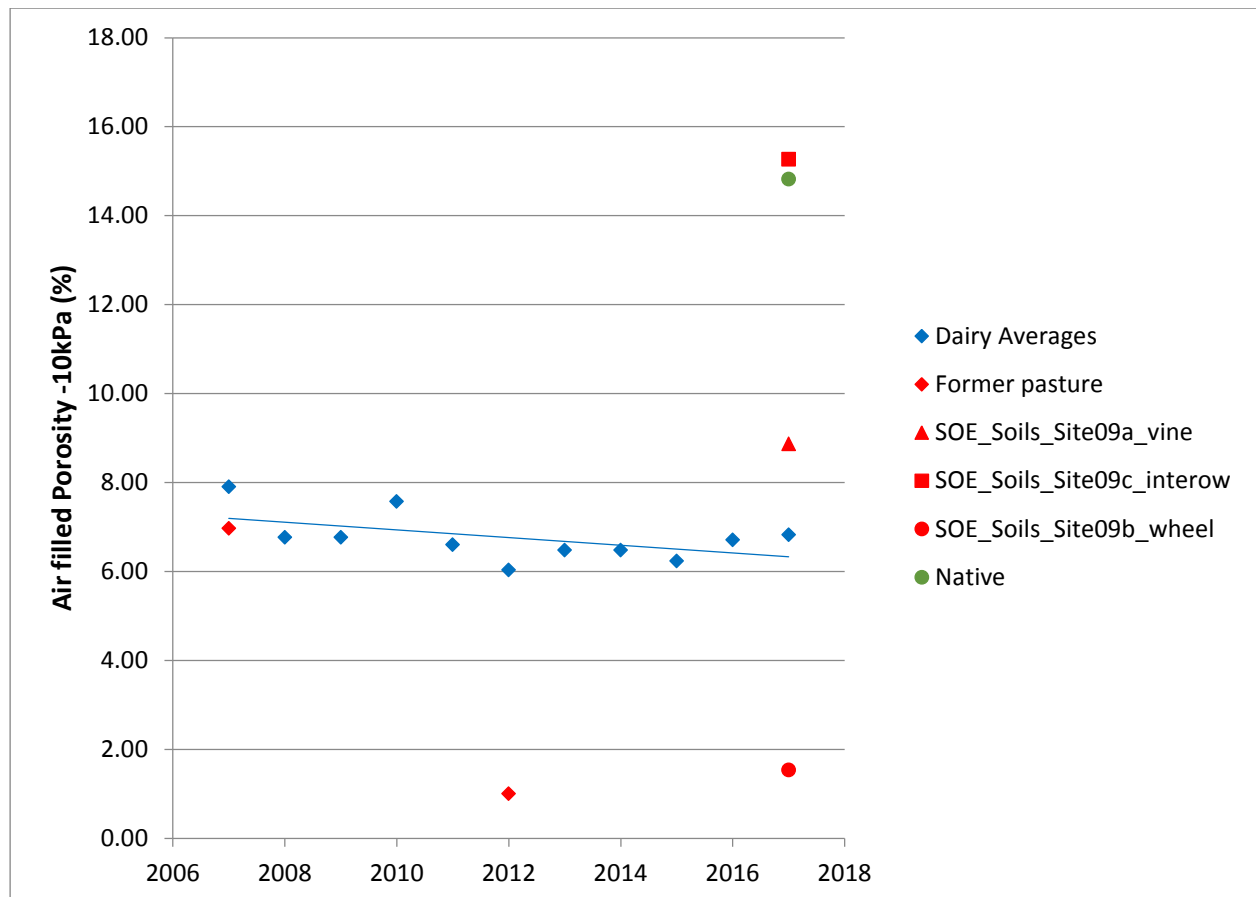


Figure 30: Site 9, Air filled porosity of former dairy pasture following conversion to viticulture

4.5.3. Land use change - Discussion

While noting the sparse data set used in this section, the speed that these changes in soil physical conditions and soil organic matter have happened is the most alarming aspect of these results. This type of land use change is occurring over large parts of Marlborough annually as the viticulture industry grows. The consequences of such large losses of soil structural integrity and soil carbon are already reasonably well understood.

Decreased structural integrity (increased bulk density and reduced AFP) will lead to reduced water holding capacity and infiltration of rainwater. This will increase stormwater velocities and amounts moving across the soil surface leading to greater inputs of sediment and nutrients into waterways (Mclaren & Cameron, 1996; Havlin et al, 2013).

Carbon loss is potentially a greater problem. Carbon is lost from the soil via the respiration of soil microbes which is stimulated by cultivation. Each tonne of carbon lost from the soil equates to 3.67 tonnes of carbon dioxide. If Site 28 is representative, then the 2.4% of total carbon lost equates to 46.8 tonnes of carbon per hectare or 171 tonnes of carbon dioxide. For perspective, a hectare of pines sequesters between 4-7 tonnes of carbon dioxide per year. Effectively, to recover the lost soil carbon from a hectare of newly established vineyard, one hectare of pines would need to grow for a full rotation of 25 years.

5. Summary

Results for the 2017 Soil Quality Monitoring round showed trends consistent with last years results. Soil compaction, soil phosphate levels and loss of organic matter are the main concerns this year. Sixty percent of sites showed low air filled porosity indicating a reduction in pore space in the soil. Soil phosphate levels were found to be outside the target ranges for 65% of samples. While only 1 site had excessive phosphate levels, most had levels below the agronomic optimal levels indicating they may be performing below optimal levels. Similar to 2016, these concerns were noted across all soil types sampled but only on farmed land. Forest sites reported both low bulk Density and high air filled porosity indicating no soil compaction.

Discussion of long-term trends introduced in 2016 continues in this report. The same issues are still of most concern, these being the risk of nutrient loss to water, soil compaction and loss of organic matter. A new section has been introduced this year that seeks to quantify the changes that conversion from pastoral land uses to viticulture brings. Although this is a very small data set, soil quality changes can be seen very quickly after conversion. These changes include large falls in total carbon, increases in bulk density and decreases in air filled porosity. These all relate directly to the extensive soil cultivation that occurs at vineyard establishment and the consequent vineyard management practices.

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8. Appendix A. Soil Target Values

Soil quality indicator target (or optimal) ranges from Hill and Sparling (2009) are outlined in the tables below along with guideline values for trace element concentrations in soil, adapted from NZWWA (2003). Olsen P values as set by Taylor and Mackay (2012).

Bulk density target ranges (t/m³ or Mg/m³)

	Very loose	Loose	Adequate	Compact	Very compact	
Semi-arid, Pallic and Recent soils	0.3	0.4	0.9	1.25	1.4	1.6
Allophanic soils		0.3	0.6	0.9	1.3	
Organic soils		0.2	0.4	0.6	1.0	
All other soils	0.3	0.7	0.8	1.2	1.4	1.6

Notes:

Applicable to all land uses

Target ranges for cropping and horticulture are poorly defined

Air filled porosity target ranges (% @ -10 kPa)

	Very low	Low	Adequate	High	
Pastures, cropping and horticulture	0	6	10¹	30	40
Forestry	0	8	10	30	40

Notes:

1: Revised based on Mackay et al. (2006)

Applicable to all Soil Orders

Target ranges for cropping and horticulture are poorly defined

Total carbon target ranges (% w/w)

	Very depleted	Depleted	Normal	Ample	
Allophanic	0.5	3	4	9	12
Semi-arid, Pallic and Recent	0	2	3	5	12
Organic	exclusion				
All other Soil Orders	0.5	2.5	3.5	7	12

Notes:

Applicable to all Soil Orders

Organic soils by definition must have >15% total C content, hence C content is not a quality indicator for that order and is defined as an "exclusion" Target ranges for cropping and horticulture are poorly defined

Total nitrogen target ranges (% w/w)

	Very depleted	Depleted	Normal	Ample	High	
Pasture	0	0.25	0.35	0.65	0.70	1.0
Forestry	0	0.10	0.20	0.60	0.70	
Cropping and horticulture	exclusion					

Notes:

Applicable to all Soil Orders

Target ranges for cropping and horticulture are not specified as target values will depend on the specific crop grown

Anaerobic mineralisable nitrogen (AMN) target ranges (mg/kg)

	Very low	Low	Adequate	Ample	High	Excessive	
Pasture	25	50	100	200	200	250	300
Forestry	5	20	40	120	150	175	200
Cropping and horticulture	5	20	100	150	150	200	225

Notes:

Applicable to all Soil Orders

Target ranges for cropping and horticulture are poorly defined

Soil pH target ranges

	Very acid	Slightly acid	Optimal	Sub-optimal	Very alkaline	
Pastures on all soils except Organic	4	5	5.5	6.3	6.6	8.5
Pastures on Organic soils	4	4.5	5	6	7.0	
Cropping and horticulture on all soils except Organic	4	5	5.5	7.2	7.6	8.5
Cropping and horticulture on Organic soils	4	4.5	5	7	7.6	
Forestry on all soils except Organic		3.5	4	7	7.6	
Forestry on Organic soils	exclusion					

Notes:

Applicable to all Soil Orders

Target ranges for cropping and horticulture are general averages and target values will depend on the specific crop grown

Exclusion is given for forestry on organic soils as this combination is unlikely because of wind throw

Olsen P target ranges (mg/L or $\mu\text{g}/\text{cm}^3$)

	Very low	Low	Adequate	High	
Pasture on Sedimentary and Allophanic soils	0	15	20	50	200
Pasture on Pumice and Organic soils	0	15	35	50	200
Cropping and horticulture on Sedimentary and Allophanic soils	0	20	50	50	200
Cropping and horticulture on Pumice and Organic soils	0	25	60	50	200
Forestry on all Soil Orders	0	5	10	50	200

Notes:

Sedimentary soil includes all other Soil Orders except Allophanic (volcanic ash), Pumice, Organic and Recent (AgResearch classification system)

Guideline values for trace element concentrations in soil, adapted from NZWWWA (2003)

Trace element	Soil Limit (mg/kg)
Arsenic (As)	20
Cadmium (Cd)	1
Chromium (Cr)	600
Copper (Cu)	100
Lead (Pb)	300
Nickel (Ni)	60
Zinc (Zn)	300